

Article

Nonpeptidic Z360-Analogs Tagged with Trivalent Radiometals as Anti-CCK₂R Cancer Theranostic Agents: A Preclinical Study

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Abstract: (1) Background: Theranostic approaches in the management of cholecystokinin subtype 2 receptor (CCK₂R)-positive tumors include radiolabeled gastrin and CCK motifs. Moving toward antagonist-based CCK₂R-radioligands instead, we herein present three analogs of the nonpeptidic CCK₂R-antagonist Z360, GAS1/2/3. Each was conjugated to a different chelator (DOTA, NODAGA or DOTAGA) for labeling with medically relevant trivalent radiometals (e.g., Ga-68, In-111, Lu-177) for potential use as anti-CCK₂R cancer agents; (2) Methods: The in vitro properties of the three analogs were compared in stably transfected HEK293-CCK₂R cells. Biodistribution profiles were compared in SCID mice bearing twin HEK293-CCK₂R and wtHEK293 tumors; (3) Results: The GAS1/2/3 analogs displayed high CCK₂R-affinity (lower nM-range). The radioligands were fairly stable in vivo and selectively targeted the HEK293-CCK₂R, but not the CCK₂R-negative wtHEK293 tumors in mice. Their overall pharmacokinetic profile was found strongly dependent on the radiometal-chelate. Results could be visualized by SPECT/CT for the [¹¹¹In]In-analogs; (4) Conclusions: The present study highlighted the high impact of the radiometal-chelate on the end-pharmacokinetics of a new series of Z360-based radioligands, revealing candidates with promising properties for clinical translation. It also provided the impetus for the development of a new class of nonpeptidic radioligands for CCK₂R-targeted theranostics of human cancer.



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Keywords: tumor theranostics; CCK₂R-radioligand; Z360; nonpeptidic radioligand; SPECT; PET; radionuclide therapy; [¹¹¹In]In/[^{67/68}Ga]Ga/[¹⁷⁷Lu]Lu-radioligand

1. Introduction

The cholecystokinin subtype 2 receptor (CCK₂R) represents a clinically relevant biomolecular target for nuclear medicine applications owing to its overexpression in a variety of human tumors, such as medullary thyroid carcinoma (MTC), small cell lung cancer (SCLC), astrocytomas, stromal ovarian cancers or gastrointestinal stromal tumors (GIST) [1–6]. In view of that, a good number of peptide radioligands were previously developed with the aim to deliver diagnostic or therapeutic radionuclides specifically on CCK₂R-positive cancer lesions [7,8]. Thus far, the native peptides cholecystokinin (e.g., sulfated CCK-8, H-Asp-Tyr(SO₃H)-Met-Gly-Trp-Met-Asp-Phe-NH₂) and gastrin (gastrin-17, pGlu-Gly-Pro-Trp-Leu-Glu-Glu-Glu-Glu-Ala-Tyr-Gly-Trp-Met-Asp-Phe-NH₂) have served as motifs in anti-CCK₂R radioligand design. The coupling of suitable chelators has enabled labeling with radiometals clinically applied for diagnosis and therapy—“theranostics”. Following this concept, diagnostic imaging with single-photon emission tomography (SPECT:

In-111) or positron emission tomography (PET: Ga-68) serves to select patients eligible for radionuclide therapy with particle emitters (e.g., Lu-177, Y-90 or Ac-225). Targeted radiotherapy aims at the eradication of tumor cells only, sparing healthy tissues devoid of CCK₂R-expression. Thereafter, imaging is applied again to monitor therapeutic responses and disease progression in a patient-tailored protocol.

A major challenge in the development of clinically useful CCK₂R-radioligands is the fast in vivo degradation of truncated *des*-(Glu)₅ gastrin and minigastrin (MG: gastrin(4–17)) analogs [9–14]. Notably, the presence of the (Glu)₅-chain, although shown to favor metabolic stability and tumor targeting of related radioligands, led to prolonged kidney retention. In a smart, recent approach, the administration of key protease inhibitors successfully stabilized *des*-(Glu)₅-gastrin radioligands in vivo, significantly enhancing CCK₂R-positive tumor uptake in mice and MTC patients [15,16]. In another approach, innovative structural interventions on the peptide chain led to metabolic stability improvements without compromising other important properties of resulting radioligands [17–21]. The clinical value of the selected new analogs is currently being assessed through pilot studies in MTC patients [22–24]. Thus far, all tested radioligands have been peptide-based, displaying agonistic properties at the CCK₂R. Consequently, they are prone to elicit adverse effects after binding and activation of the CCK₂R when injected to patients. These effects are well described during the provocative pentagastrin (Boc-βAla-Trp-Met-Asp-Phe-NH₂) test used in the early diagnosis and follow up of MTC [25]. Another point of concern is the tendency of radiolabeled gastrin analogs to accumulate in the gastric mucosa, a tissue of high physiological CCK₂R-expression levels, a feature harboring dosimetric restrictions, especially during radiotherapy [17,26].

It is reasonable to assume that such concerns can be efficiently addressed by the use of radiolabeled CCK₂R-antagonists, based on the recent positive experience from the fields of somatostatin and bombesin radioligands. It should be noted that in the case of CCK₂R, the most known antagonists considered as motifs in radioligand design are non-peptidic small organic molecules [27]. Benzodiazepines represent a major class thereof, with Z360 (3-[[1-cyclohexyl-5-(3,3-dimethyl-2-oxobutyl)-4-oxo-2,3-dihydro-1,5-benzo-diazepin-3-yl]-carbamoylamino-benzoic acid) in particular having recently attracted our attention. Z360 was first reported as a CCK₂R-antagonist, shown to inhibit the meal- or pentagastrin-induced acid secretion in the gastric mucosa in dogs after systemic administration [28,29]. Next, Z360 was shown to play a role in several gastrointestinal pathological processes, including cancer, and was eventually proposed as a drug against pancreatic cancer [30–34].

The feasibility of attaching functional groups, such as cytotoxic drugs, dyes or metal-chelates, at the free carboxylic acid of Z360, without compromising the affinity of resulting bioconjugates for the CCK₂R, was recently established [35–37]. We previously reported on [^{99m}Tc]Tc-DGA1 (DGA1: Z360-PEG₃-DGlu-DGlu-DGlu-DGu-Lys(N₄)-OH; N₄: 6-carboxy-1,4,8,11-tetraazaundecane), our first Z360 [^{99m}Tc]Tc-based radioligand, suitable for imaging CCK₂R-positive tumors with SPECT and SPECT/CT [38]. Aiming to broaden the applicability of Z360 in PET imaging and radionuclide therapy, we now present three new analogs, GAS1/2/3 (Figure 1), amenable for labeling with theranostic trivalent radiometals. For this purpose, the macrocyclic chelators, DOTA (1,4,7,10-tetraazacyclododecane-1,4,7,10-tetraacetic acid), NODAGA (1,4,7-triazacyclononane,1-glutaric acid-4,7-acetic acid) or DOTAGA (1,4,7,10-tetraazacyclododecane,1-(glutaric acid)-4,7,10-triacetic acid) were coupled at the pendant carboxylate group of Z360 via the common H-DGlu-HN(PEG₃)NH₂ (α,ω-diamino tri-(ethylene glycol)) linker. This arrangement has allowed for the direct comparison of biological responses across analogs after labeling them with representative radiometals used in cancer theranostics, specifically for PET (Ga-67 as a surrogate of Ga-68), SPECT (In-111) and radionuclide therapy (Lu-177). Accordingly, pharmacological features of resulting analogs could be directly compared in CCK₂R-positive cell preparations and their in vivo profiles in mice models. In this way, radioligands were screened to select candidates for further clinical evaluation. Moreover, conclusions on the strengths and

limitations of using Z360-based radioligands, and possibly CCK₂R-antagonists in general, in cancer theranostics could be drawn.

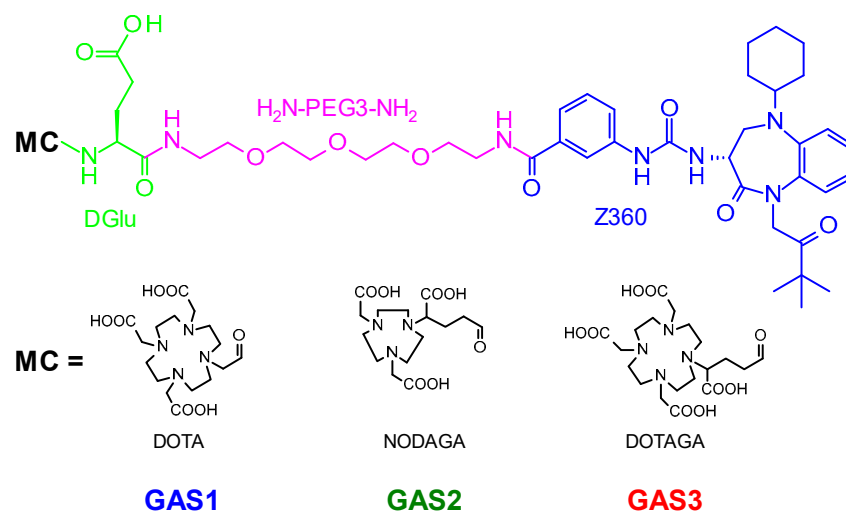


Figure 1. Chemical structure of GAS1, GAS2 and GAS3.

2. Materials and Methods

2.1. Chemicals and Ligands

Z360 was purchased from MedKoo Biosciences, Inc. (Morrisville, NC/USA). The three Z360-conjugates (Figure 1): (i) GAS1, DOTA-DGLu-HN(PEG3)NH-Z360, (ii) GAS2, NODAGA-DGLu-HN(PEG3)NH-Z360 and (iii) GAS3, DOTAGA-DGLu-HN(PEG3)NH-Z360, as well as DG2 were synthesized by PiChem Forschungs- und Entwicklungs GmbH (Raaba-Grambach, Austria); analytical data, comprising MALDI-TOF mass spectroscopy results and results from HPLC analysis in two distinct HPLC systems, is compiled in Table S1 (Supplementary File). The human [Leu¹⁵]gastrin-17 (pGlu-Gly-Pro-Trp-Leu-(Glu)₅-Ala-Tyr-Gly-Trp-Leu-Asp-Phe-NH₂) was purchased from Bachem (Bubendorf, Switzerland). For metal incorporation by the GAS-analogs, the nitrate salts [^{nat}Ga]Ga(NO₃)₃, [^{nat}In]In(NO₃)₃ and [^{nat}Lu]Lu(NO₃)₃ were obtained from Sigma-Aldrich. Gelofusine was obtained from B. Braun Melsungen AG (Melsungen, Germany).

2.2. Labeling of GAS-Compounds with Ga-67, In-111 and Lu-177

All three GAS analogs were labeled with Ga-67 (serving as a surrogate of the PET radionuclide Ga-68) and In-111, but only GAS1 and GAS3 were labeled with Lu-177. Lyophilized GAS1/2/3 were dissolved in HPLC-grade H₂O (2 mg/mL), and 50 µL aliquots thereof were stored in Eppendorf Protein LoBind tubes at −20 °C. For Ga-67 labeling, [⁶⁷Ga]GaCl₃ (4–5.5 GBq/mL in dilute HCl solution) was provided by IDB Holland B.V. (Baarle-Nassau, The Netherlands). For labeling with In-111, [¹¹¹In]InCl₃ (400–600 MBq/mL in 0.05 mM HCl) was purchased from Curium Pharma (Petten, The Netherlands). For labeling with Lu-177 [¹⁷⁷Lu]LuCl₃ (3.7 GBq/mL in 0.04 M HCl, RLu-3, A_s > 370 GBq/mg Lu) were obtained from POLATOM (Otwock, Poland). Labeling protocols for each radionuclide are presented in detail in the Supplementary File.

For the preparation of [¹²⁵I][I-Tyr¹²,Leu¹⁵]gastrin-17, [¹²⁵I]NaI in a 0.1 M NaOH solution (Perkin Elmer) was employed, as previously described [38]. Aliquots of the radioligand stock solution in 0.1% BSA-PBS buffer were kept at −20 °C and were used in competition binding experiments (molecular activity of 74 GBq/µmol). The preparation of [^{nat}In]In/[^{nat}Ga]Ga-GAS1/2/3 and [^{nat}Lu]Lu-GAS1/3 is detailed in the Supplementary File.

2.3. Quality Control of Radiolabeled GAS1/2/3

Reversed-phase high-performance liquid chromatography (RP-HPLC) was performed on a Waters Chromatograph based on a 600 E multi-solvent delivery system coupled to

a Waters 2998 photodiode array detector (Waters, Vienna, Austria) and a Gabi gamma-detector (Raytest, RSM Analytische Instrumente GmbH, Straubenhardt, Germany). The processing of data and chromatography were controlled by Empower Software (Waters, Milford, MA, USA). For quality control, aliquots of the radiolabeling solution were loaded on a Symmetry Shield RP18 cartridge column (5 μm , 3.9 mm \times 150 mm, Waters, Eschborn, Germany), eluted with the following linear gradient: 100%A/0% B to 70%A/30% B in 5 min and then 70%A/30% B to 55%A/45% B in 60 min, whereby A = 0.1% TFA in H₂O (*v/v*) and B = MeCN (system 1). The radiochemical labeling yields exceeded 98%, and the radiochemical purity was >99%; therefore, radioligands were used without further purification in all subsequent experiments. Samples of [¹¹¹In]In/[⁶⁷Ga]Ga-GAS1/2/3 and [¹⁷⁷Lu]Lu-GAS1/3 were tested before and after the end of all biological experiments.

Handling of solutions containing beta-/gamma-emitting radionuclides was conducted by authorized personnel in compliance with European radiation safety guidelines. Licensed facilities were supervised by the Greek Atomic Energy Commission (GAEC, license #A/435/17092/2019 and #A/435/15767/2019).

2.4. Cell Culture

For most biological experiments, HEK293 cells transfected to stably express the human CCK_{2i4sv}R splice variant were used, kindly provided by Dr. P. Laverman (Radboud University Nijmegen Medical Center, Nijmegen, The Netherlands) and Prof. M. R. Hellmich (University of Texas Medical Branch, Galveston, TX, USA) [39–41], while non-transfected wtHEK293 cells served as negative controls. Cells were cultured in DMEM with GlutaMAX-I supplemented with 10% (*v/v*) fetal bovine serum (FBS), 100 U/mL penicillin and 100 $\mu\text{g}/\text{mL}$ streptomycin and kept in a controlled humidified air containing 5% CO₂ at 37 °C; the medium for transfected cells was additionally supplemented with 400 $\mu\text{g}/\text{mL}$ G418. All culture media were purchased from Gibco BRL, Life Technologies (Grand Island, NY, USA) and supplements were supplied by Biochrom KG Seromed (Berlin, Germany). For Ca²⁺-mobilization assays, HEK293 cell clones (generated by O.G. Chepurny in the G.G. Holz laboratory) were transfected to stably co-express either both the human CCK₂R and the Ca²⁺ biosensor YC3.60 (clone #21) or only YC3.60 (clone #5, negative controls devoid of CCK₂R expression) (Supplementary File). Ca²⁺ mobilization assays were conducted by FRET-based detection of Ca²⁺ in HEK293-hCCK₂R/YC3.60 and HEK293-YC3.60 cells (negative controls) for GAS1/2/3 as well as for DG2, [Leu¹⁵]gastrin-17 and Z360 (controls), as outlined in the Supplementary File.

2.5. Competition Binding Assays in HEK293-CCK_{2i4sv}R Cell Membranes

Competition binding assays for the metal-free GAS1/2/3 and the corresponding metal-tagged [^{nat}Ga]Ga/[^{nat}In]In/[^{nat}Lu]Lu-GAS1/3 and [^{nat}Ga]Ga/[^{nat}In]In-GAS2 analogs were conducted against the [¹²⁵I][I-Tyr¹²,Leu¹⁵]gastrin-17 radioligand in membrane homogenates harvested from HEK293-CCK_{2i4sv}R cells, as previously described [38,42]. Briefly, each of the above analogs, or unmodified Z360 serving as control, was incubated in triplicates of increasing concentrations (10⁻¹³–10⁻⁶ M) with [¹²⁵I][I-Tyr¹²,Leu¹⁵]gastrin-17 (50 pM, \approx 40,000 cpm) and membrane homogenate in binding buffer (300 μL , pH 7.4, 50 mM HEPES, 1% BSA, 5.5 mM MgCl₂, 35 μM bacitracin) for 1 h at 22 °C. Incubation was interrupted by rapid filtration through glass fiber filters (Whatman GF/B, presoaked in binding buffer for at least 1 h) on a Brandel Cell Harvester (Adi Hassel Ingenieur Büro, Munich, Germany) and subsequent rinsing with ice-cold washing buffer (10 mM HEPES pH 7.4, 150 mM NaCl). The radioactivity content of individual filters was measured in a gamma counter (automated multi-sample well-type instrument with a NaI(Tl) 3' crystal, Canberra Packard Cobra™ Quantum U5003/1, Auto-Gamma® counting system). The 50% inhibitory concentration (IC₅₀) was determined by nonlinear regression analysis according to a one-site model using PRISM 6 (Graph Pad Software, San Diego, CA, USA). Values are expressed as mean \pm standard deviation (SD) of three independent experiments performed in triplicate.

2.6. Uptake of Radiolabeled GAS1/2/3 in HEK293-CCK_{2i4sv}R Cells

A day before the experiment, HEK293-CCK_{2i4sv}R cells were seeded in six-well plates. The next day, cells were rinsed with ice-cold internalization medium (IM: DMEM Glutamax-I, supplemented by 1% (*v/v*) FBS) and then the fresh medium was added (1.2 mL) at 37 °C. A further portion of IM (150 µL) was added in the upper well-row, and DG2 solution in IM (150 µL) was added in the lower row (to a final concentration of 1 µM; non-specific series). Each of the test radioligands, [⁶⁷Ga]Ga/[¹¹¹In]In/[¹⁷⁷Lu]Lu-GAS1/3 and [⁶⁷Ga]Ga/[¹¹¹In]In-GAS2, was finally added (250 fmol total conjugates in 150 µL 0.5% BSA-PBS) in all the wells and the plates were incubated at for 1 h at 37 °C in an Incubator-Orbital Shaker unit (MPM Instr. Srl, Bernareggio, MI, Italy); [¹¹¹In]In-GAS1/2/3 were incubated for 15 min, 30 min, 1 h and 2 h. At the predetermined time points, the plates were placed on ice, the medium was collected, and the plates were rinsed with 0.5% BSA-PBS (1 mL). Cells were subsequently treated with an acid-wash solution (2 × 600 µL; 50 mM glycine buffer pH 2.8, 0.1 M NaCl) and the fractions collected (membrane-bound). After briefly washing with 0.5% BSA-PBS (1 mL), the cells were lysed with 1 N NaOH (2 × 600 µL), and the fractions were collected (internalized). The radioactivity content of collected fractions was counted in the gamma counter, and the percentage of internalized and membrane-bound fractions per time point were calculated with Microsoft Excel. Specific internalized and membrane-bound counts were determined by subtracting the respective non-specific (in the presence of 1 µM DG2) from the respective total counts. Results represent specific internalized/membrane bound ± SD of total added radioactivity per well from three experiments performed in triplicate.

2.7. Radioligand Stability in Mice

The formation of radiometabolites which can be detected in the blood was tested in blood samples collected 5 min post-injection (pi) of each of the radioligands [⁶⁷Ga]Ga/[¹¹¹In]In/[¹⁷⁷Lu]Lu-GAS1/3 and [⁶⁷Ga]Ga/[¹¹¹In]In-GAS2. For this study, in-house male Swiss albino mice (body weight: 30 ± 5 g, provided by the NCSR “Demokritos” Animal House) were used in groups of three. The animals received, through the tail vein, a 100-µL bolus containing each radioligand tested (2.5–3 nmol of the total conjugate in vehicle: saline/EtOH 9/1 *v/v*, corresponding to: up to 11 MBq for In-111, up to 13 MBq for Ga-67 and up to 74 MBq for Lu-177). Mice were euthanized, and blood was collected directly from the heart using a prechilled penicillin syringe and transferred in a prechilled EDTA-containing Eppendorf Protein LoBind[®] tube on ice (40 µL, 50 mM Na₂EDTA solution). After centrifugation (10 min, 2000 × *g*/4 °C, in a Hettich, Universal 320R, centrifuge), the blood plasma was collected, mixed with chilled MeCN in a 1/1 *v/v* ratio and centrifuged again (10 min, 15,000 × *g*/4 °C). Supernatants were collected and concentrated to a small volume (≈50–100 µL) under a gentle N₂-flux at 40 °C. They were then diluted with physiological saline (≈400 µL) and filtered through a Millex GV filter (0.22 µm, 13 mm Ø, Millipore, Milford, CT, USA). Aliquots of the filtrate were analyzed by radio-RP-HPLC using a Waters Symmetry Shield RP18 cartridge column (5 µm, 3.9 mm × 20 mm) eluted at a flow rate of 1.0 mL/min with the following gradient: 100% A/0% B to 55% A/45% B in 45 min; A = 0.1% aqueous TFA (*v/v*) and B = MeCN (system 2). The elution time (*t_R*) of each parent radioligand was determined by coinjection of blood samples with an aliquot of the labeling solution on the HPLC column. Results were calculated as the average percentage of intact radioligand ± SD derived from three independent experiments.

Following the same injection protocol, radioligands were administered in additional groups of mice, but this time animals were euthanized 30 min pi. The urine was immediately collected from the bladder using a prechilled syringe and transferred as before in a prechilled EDTA-containing Eppendorf Protein LoBind[®] centrifuge tube on ice (40 µL, 50 mM Na₂EDTA solution) and 0.1 mL of physiological saline. Urine proteins were precipitated by adding chilled MeCN in a 1/1 *v/v* ratio followed by a 10 min centrifugation step (15,000 × *g*/4 °C). The supernatant was then collected and treated following the blood sample workup procedure. Aliquots of the Millex-GV filtrate (containing >90% of total

urine activity) were analyzed by radio-HPLC, adopting the aforementioned conditions. Results were calculated as the average percentage of intact radioligand \pm SD derived from three independent experiments.

2.8. Biodistribution in Mice Bearing Twin HEK293-CCK_{2i4sv}R and wtHEK293 Xenografts

Inocula containing a suspension of freshly harvested HEK293-CCK_{2i4sv}R/wtHEK293 cells (150 μ L, $1.8 \times 10^7/1.4 \times 10^7$ cells, respectively, in normal saline) were subcutaneously (sc) injected in the right and left flanks of male SCID mice (24.5 ± 2.3 g body weight, six weeks of age on arrival day, NCSR “Demokritos” Animal House Facility). Well-palpable tumors ($0.42 \pm 0.37/0.25 \pm 0.32$ mg, respectively) were grown at the inoculation site 3–4 weeks later, and biodistribution was conducted. On the day of biodistribution, animals were divided into groups of four and received the radioligand through the tail vein as a bolus (100 μ L, containing each of: i. 78–91 kBq [⁶⁷Ga]GAS1/2/3 corresponding to 7–17 pmol GAS1/2/3; ii. 74–92 kBq [¹¹¹In]GAS1/2/3 corresponding to 20–25 pmol GAS1/2/3; iii. 730 kBq [¹⁷⁷Lu]Lu-GAS1/3 corresponding to 40 pmol GAS1/3, in-vehicle: saline/EtOH 9/1 *v/v*). Animals were euthanized at predetermined time intervals depending upon the radionuclide applied (for [⁶⁷Ga]Ga-GAS1/2/3: 1, 4 and 24 h pi; for [¹¹¹In]In-GAS1/2/3: 4 and 24 h pi—two extra groups of mice receiving geflofusine together with [¹¹¹In]In-GAS1/2 at 4 and 24 h pi were included; and for [¹⁷⁷Lu]Lu-GAS1/3: 4, 24, 48, 72 and 96 h pi). Mice were dissected, and blood samples, organs of interest and tumors were rapidly collected, weighted and counted in the gamma counter. Biodistribution data was calculated as the percentage of the administered dose per gram tissue (%IA/g) with the aid of suitable standards of the administered dose and the Microsoft Excel program. Results are presented as mean % IA/g values \pm SD, $n = 4$ per time point, as determined by the PRISM™ 6.0 GraphPad software (San Diego, CA, USA).

2.9. Statistical Analysis

The statistical evaluation of results was carried out by applying a two-way ANOVA with multiple comparisons and Tukey’s post hoc analysis (PRISM™ 6.0 GraphPad Software, San Diego, CA/USA). *p*-values of <0.05 were considered to be statistically significant.

2.10. SPECT/CT of HEK293-CCK_{2i4sv}R and wtHEK293 Xenografts with [¹¹¹In]In-GAS1/2/3

For SPECT/CT imaging, six mice bearing twin HEK293-CCK_{2i4sv}R and wtHEK293 tumors were injected in the tail vein with a bolus containing [¹¹¹In]In-GAS1/2/3 (100 μ L, 9–12 MBq associated with 3 nmol total injected analog in-vehicle: saline/EtOH 9/1 *v/v*). Mice were euthanized at 4 h and 24 h pi. Tomographic SPECT/CT imaging was performed with the γ -CUBE/ x -CUBE systems (Molecubes, Belgium). The SPECT system is based on monolithic NaI detectors attached to SiPMs, with a 0.6 mm intrinsic resolution. The CT system is based on a structured CMOS detector of CsI with pixels of 75 μ m and operates between 35–80 kVp, 10–500 μ A tube current, with a 33 μ m fixed focal spot size. SPECT scans were acquired 4 h pi and 24 h pi, with a 40–50 min duration protocol based on the injected activity, and each SPECT scan was succeeded by a CT scan, following General-Purpose protocol under 50 kVp, for co-registration purposes. SPECT images were reconstructed using the MLEM reconstruction method with a 250 μ m voxel size and 500 iterations. CT images were reconstructed using the ISRA reconstruction method with a 100 μ m voxel size.

Images were exported and post-processed on VivoQuant software, version 4.0 (Invicro, Boston). A smoothing median filter (0.6 mm, spherical window) was applied to the images, and the bladder was removed for consistency purposes. Normalization of images was performed (i.e., all images having the same color scale range values) to achieve a direct visual comparison between the different groups.

All experiments involving mice were conducted in compliance with European and national regulations in licensed facilities (EL 25 BIO exp021). The study protocols were approved by the Department of Agriculture and Veterinary Service of the Prefecture of Athens

(#1609, 24-04-2019 for the stability studies and #1610, 24-04-2019 for the biodistribution and imaging studies).

3. Results

3.1. Ligands and Radioligands

Analytical data for the Z360 analogs GAS1/2/3, carrying the three different chelators DOTA, NODAGA and DOTAGA via a DGlu-HN(PEG3)NH-linker (Figure 1), comprising results from HPLC analyses in two separate systems and MALDI-TOF mass spectrometry data, are summarized in Table S1 (Supplementary File) and were found to be consistent with the formation of the desired products in high purity ($\geq 95\%$).

The three bioconjugates, GAS1/2/3, were labeled with Ga-67 (as a Ga-68 surrogate), In-111 and Lu-177, following previously described protocols after a slight modification to avoid by-products caused by the degradation of Z360 at elevated temperatures. Under these precautions, the final radiolabeled products were obtained in $>98\%$ purity, as verified by the respective radio-analytical HPLC (Supplementary File).

3.2. Receptor Affinity Determination and Functional Studies of Z360-Analogs

The receptor binding affinities of the metal-free GAS1/2/3 and the corresponding metal-tagged $^{nat}\text{Ga}/^{nat}\text{In}/^{nat}\text{Lu}$ -GAS1/3 and $^{nat}\text{Ga}/^{nat}\text{In}$ -GAS2 analogs were determined via competition binding assays in HEK293-CCK_{2i4sv}R cell membrane homogenates using [¹²⁵I][I-Tyr¹²,Leu¹⁵]gastrin-17 as the radioligand and unmodified Z360 as control. All tested analogs displaced [¹²⁵I][I-Tyr¹²,Leu¹⁵]gastrin-17 from HEK293-CCK_{2i4sv}R binding sites on the membranes in a mono-phasic and concentration-dependent way.

As summarized in Table 1, the coupling of the metal-chelator led to a drop in the receptor affinities of GAS1/2/3 compared to the unmodified Z360; this drop was found to be statistically significant only in the case of the DOTA-modified analog GAS1 (IC₅₀ 5.9 ± 1.8 nM vs. IC₅₀ 1.2 ± 0.5 nM of Z360; $p < 0.05$). The binding of non-radioactive In, Ga or Lu affected the receptor affinity of the resulting metal-tagged species in different ways. Thus, no statistically significant differences could be observed in the receptor affinities between DOTAGA-modified GAS3 and its $^{nat}\text{Ga}/^{nat}\text{In}/^{nat}\text{Lu}$ -metal-tagged species. In contrast, in GAS1 (IC₅₀ 5.9 ± 1.8 nM), the incorporation of Ga (IC₅₀ 13.8 ± 2.7 nM; $p < 0.0001$) or Lu (IC₅₀ 12.4 ± 0.2 nM; $p < 0.0001$) led to a clear decline of receptor affinity. Notably, the most drastic drop of receptor affinity was displayed after the incorporation of In by the NODAGA-modified analog GAS2 (53.2 ± 2.7 nM vs. IC₅₀ 4.1 ± 1.1 nM of GAS2; $p < 0.0001$). In fact, ^{nat}In -GAS2 showed, by far, the lowest affinity for the CCK_{2i4sv}R amongst this series of Z360 analogs ($p < 0.0001$).

Table 1. Receptor affinities Z360 and its analogs as determined during competition assays against [¹²⁵I][I-Tyr¹²,Leu¹⁵]gastrin-17 in HEK293-CCK_{2i4sv}R cell membranes.

Compound	Z360	GAS1	GAS2	GAS3
metal-free	1.2 ± 0.5 (5)	5.9 ± 1.8 (3)	4.1 ± 1.1 (3)	3.1 ± 1.5 (3)
^{nat} Ga	na ¹	13.8 ± 2.7 (3)	8.8 ± 1.6 (3)	4.8 ± 0.6 (3)
^{nat} In	na ¹	5.0 ± 1.1 (3)	53.2 ± 2.7 (3)	1.9 ± 0.2 (3)
^{nat} Lu	na ¹	12.4 ± 0.2 (3)	na ¹	4.3 ± 0.9 (3)

¹ non-applicable; results are expressed in nM and represent mean IC₅₀ values ± SD with the number of experiments shown in parentheses.

Results from Ca²⁺ mobilization assays of Z360, [Leu¹⁵]gastrin-17 and DG2 (reference compounds) and GAS1/2/3 are included in Figures S1 and S2, respectively (Supplementary File). Interestingly, in the HEK293-hCCK_{2R}/YC3.60 cells, GAS1/2/3 and Z360 displayed agonistic properties only above the 100 nM concentration threshold, while the two reference agonists, [Leu¹⁵]gastrin-17 and DG2, activated the CCK_{2R} at 100 pM and 10 pM, respectively. None of the compounds triggered a response in the HEK293-YC3.60 cells, which were devoid of CCK_{2R} expression, thereby confirming the receptor-specificity of the assay.

3.3. Uptake of GAS1/2/3-Radioligands in HEK293-CCK_{2i4sv}R Cells

Comparative radioligand uptake and internalization results in HEK293-CCK_{2i4sv}R cells during 1 h incubation at 37 °C are summarized for [⁶⁷Ga]Ga-GAS1/2/3 and [¹⁷⁷Lu]Lu-GAS1/3 in Figure 2a,b, respectively; the time-dependent uptake and internalization of [¹¹¹In]In-GAS1/2/3 for the 5, 15, 30 min, 1 and 2 h incubation in the same cells are included in Figure 2c.

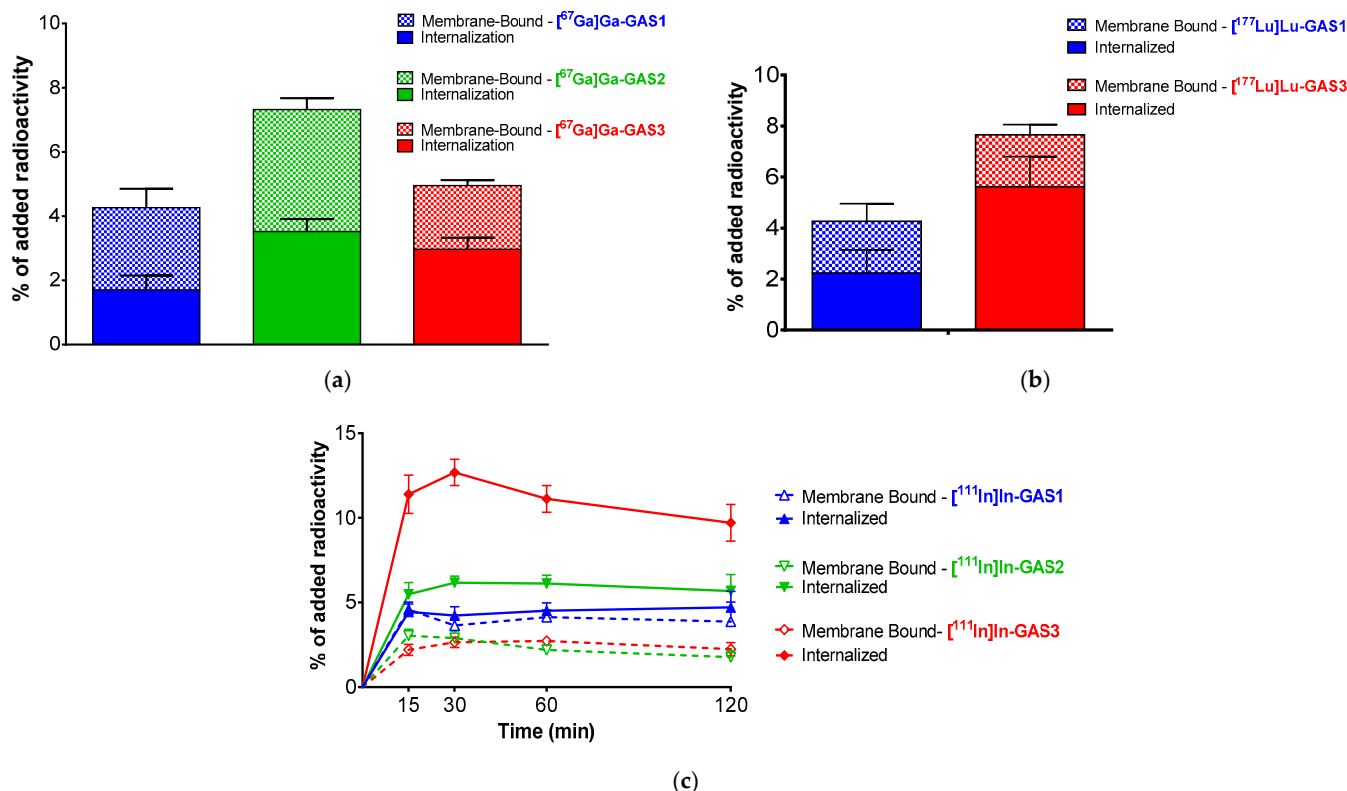


Figure 2. Radioligand uptake-internalization in HEK293-CCK_{2i4sv}R cells. (a) [⁶⁷Ga]Ga-GAS1/2/3 during 1 h incubation at 37 °C; (b) [¹⁷⁷Lu]Lu-GAS1/3 during 1 h incubation at 37 °C; (c) [¹¹¹In]In-GAS1/2/3 during 5, 15, 30 min, 1 and 2 h incubation at 37 °C. Solid bars/lines correspond to internalized fractions and chequered bars/lines to membrane-bound fractions. Results represent the average of 3 independent experiments performed in triplicate.

All analogs displayed receptor-mediated uptake by HEK293-CCK_{2i4sv}R cells with a considerable fraction of cell-associated activity found within the cells, ranging between $1.79 \pm 0.45\%$ of total added activity for [⁶⁷Ga]Ga-GAS1 to $11.12 \pm 0.79\%$ for [¹¹¹In]In-GAS3. The overall cell uptake of the radioligands was found to differ across attached chelators and radiometals. For example, within the sub-group of [⁶⁷Ga]Ga-radioligands, the cell uptake of [⁶⁷Ga]Ga-GAS2 ($7.32 \pm 0.57\%$) was found significantly higher to that of [⁶⁷Ga]Ga-GAS1 ($4.26 \pm 0.99\%$; $p < 0.0001$) or [⁶⁷Ga]Ga-GAS3 ($4.95 \pm 0.51\%$; $p < 0.0001$). On the other hand, (radio)metal incorporation was found to affect cell uptake as well. For example, in the case of GAS3 analogs, cell uptake was found to increase from [⁶⁷Ga]Ga-GAS3 ($4.95 \pm 0.51\%$) to [¹⁷⁷Lu]Lu-GAS3 ($7.65 \pm 1.31\%$; $p < 0.0001$) and [¹¹¹In]In-GAS3 ($13.81 \pm 0.89\%$; $p < 0.0001$), reaching the maximum value within the whole set of eight radioligands tested herein.

3.4. Radioligand Stability in Mice

The in vivo degradation of [⁶⁷Ga]Ga/[¹¹¹In]In/[¹⁷⁷Lu]Lu-GAS1/3 and [⁶⁷Ga]Ga/[¹¹¹In]In-GAS2 was studied via the radiometric HPLC analysis of blood samples collected 5 min pi and urine samples collected 30 min pi in healthy mice. Representative radiochromatograms of blood and urine analysis are presented in Figure 3a–f, respectively, corresponding to [⁶⁷Ga]Ga/[¹¹¹In]In/[¹⁷⁷Lu]Lu-radioligands. Numerical values are listed in Table 2.

All radioligands displayed high stability in circulation (>80%) at 5 min pi and were excreted in the urine of mice without any further decomposition at 30 min pi. The only radioligand showing significantly lower stability in the blood ($73.1 \pm 1.8\%$ intact at 5 min pi) and the urine ($44.1 \pm 2.6\%$ intact) turned out to be [^{111}In]In-GAS2, modified with the NODAGA chelator. In view of these findings, the action of peptidase inhibitors known to prolong the stability of peptide-based radioligands (e.g., the neprilysin inhibitors phosphoramidon, thiorphan or sacubitrilat and/or the angiotensin-converting enzyme inhibitor lisinopril) on the stability and biodistribution of the GAS1/2/3 radioligands were not investigated in the present study [9,12,15,43].

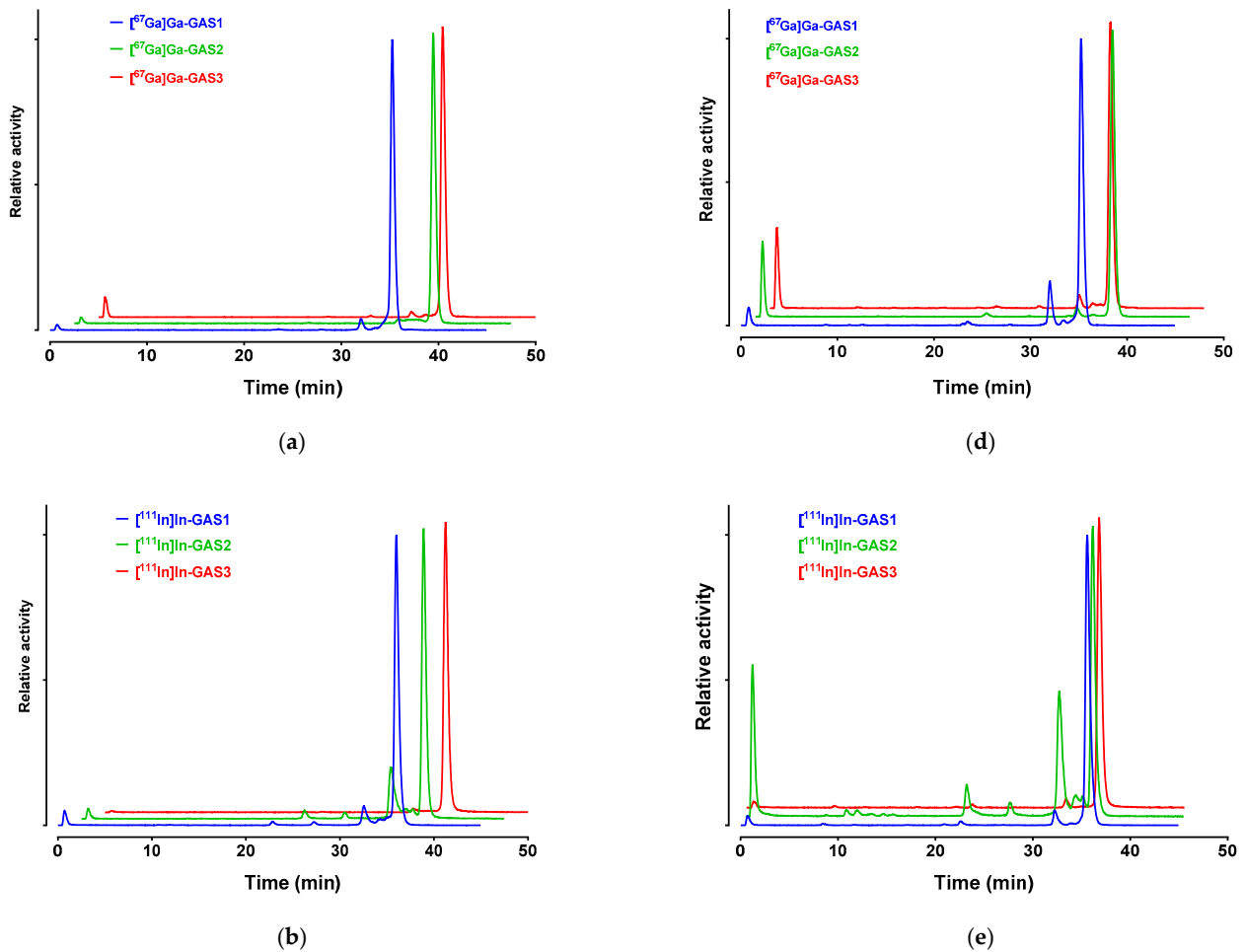


Figure 3. Cont.

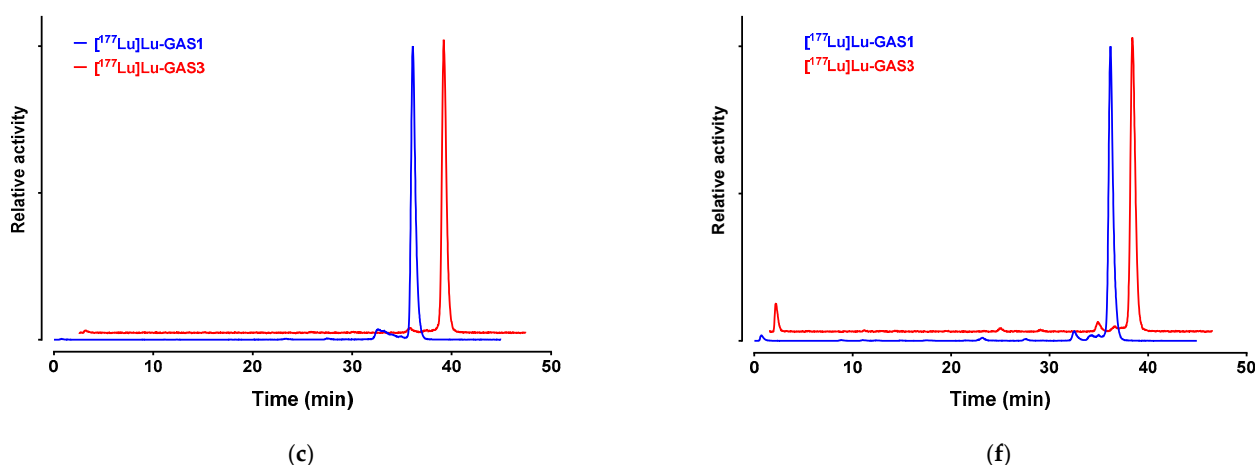


Figure 3. Representative radiochromatograms (system 2) revealing radioligand degradation in mice blood collected 5 min pi for (a) $[^{67}\text{Ga}]\text{Ga-GAS1/2/3}$, (b) $[^{111}\text{In}]\text{In-GAS1/2/3}$ and (c) $[^{177}\text{Lu}]\text{Lu-GAS1/3}$ and in urine collected 30 min pi for (d) $[^{67}\text{Ga}]\text{Ga-GAS1/2/3}$, (e) $[^{111}\text{In}]\text{In-GAS1/2/3}$ and (f) $[^{177}\text{Lu}]\text{Lu-GAS1/3}$.

Table 2. In vivo degradation of radioligands in mice blood and urine.

Radiometal Chelate/Analog		GAS1	GAS2	GAS2
$[^{67}\text{Ga}]\text{Ga}$	Blood ¹	93.9 ± 0.6 (3)	96.3 ± 3.7 (3)	92.7 ± 0.7 (3)
	Urine ²	80.2 ± 1.0 (3)	81.6 ± 1.5 (3)	78.3 ± 1.5 (3)
$[^{111}\text{In}]\text{In}$	Blood	84.6 ± 2.4 (3)	73.1 ± 1.8 (3)	98.3 ± 0.3 (3)
	Urine	85.2 ± 5.2 (3)	44.1 ± 2.6 (3)	94.3 ± 1.3 (3)
$[^{177}\text{Lu}]\text{Lu}$	Blood	81.1 ± 8.2 (3)	-	95.8 ± 2.0 (3)
	Urine	79.2 ± 10.3 (3)	-	88.1 ± 0.4 (3)

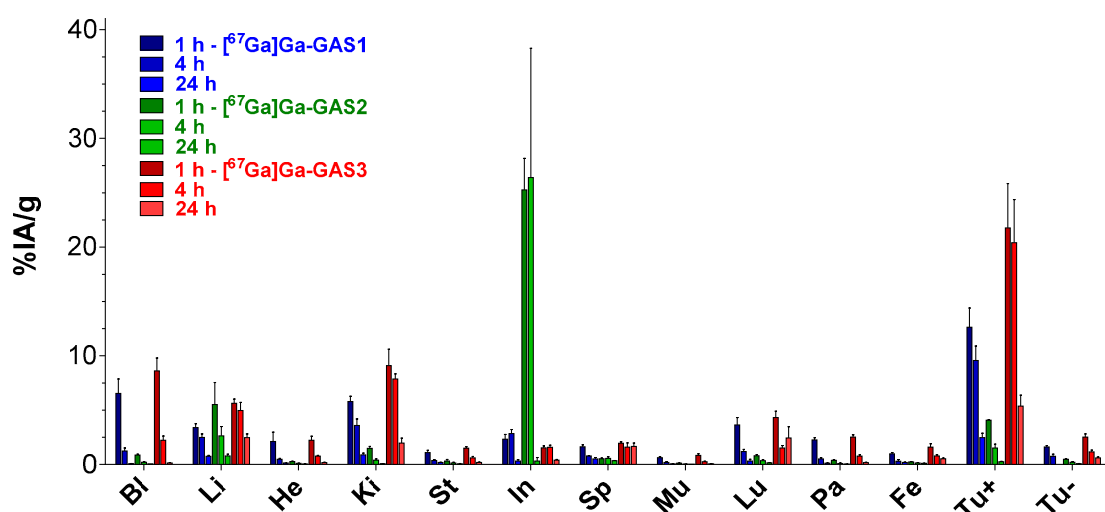
¹ Blood was collected 5 min pi; ² Urine was collected 30 min pi; results were expressed as a percentage of intact radioligand and represent mean values ± SD, with the number of independent experiments shown in parentheses.

3.5. Biodistribution in Mice Bearing Twin HEK293-CCK_{2i4sv}R and wtHEK293 Xenografts

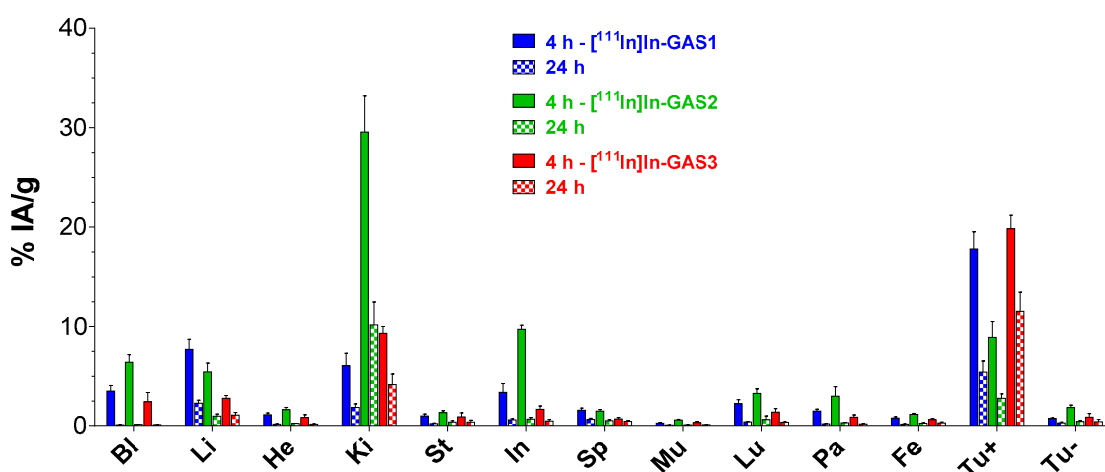
Biodistribution results for GAS1/2/3 labeled with Ga-67, In-111 and GAS1/3 labeled with Lu-177 in SCID mice bearing double HEK293-CCK_{2i4sv}R and wtHEK293 subcutaneous tumors are summarized in Figure 4a–c, respectively. Data are expressed as %IA/g and represents average values ± SD, $n = 4$ per animal group. Results in numerical values are separately displayed in Tables S2–S9 in the Supplementary File. In all cases, a notable uptake of the radioligands is evident only in the HEK293-CCK_{2i4sv}R tumors, but not in the wtHEK293 tumors, which were devoid of CCK₂R-expression, implying a receptor-mediated process. Uptake in the CCK₂R-rich stomach was found to be very low and to rapidly decline for all compounds [4]. On the other hand, the background clearance and excretion route differed amongst analogs and radiometals. It is evident that the pendant (radio)metal-chelate has a strong impact not only on receptor affinities and cell uptake but, most importantly, on the tumor-targeting capabilities and overall pharmacokinetic profile of tested GAS1/2/3-radioligands.

Specifically, in the Ga-67 set of compounds, a rank could be established with regards to uptake in the HEK293-CCK_{2i4sv}R tumors at all time intervals. For example, for the 1 h pi interval radioligands could be ranked as follows: $[^{67}\text{Ga}]\text{Ga-GAS3}$ ($21.75 \pm 4.90\% \text{IA/g}$) > $[^{67}\text{Ga}]\text{Ga-GAS1}$ ($12.62 \pm 1.76\% \text{IA/g}$; $p < 0.0001$) > $[^{67}\text{Ga}]\text{Ga-GAS2}$ ($4.06 \pm 0.07\% \text{IA/g}$; $p < 0.0001$). Of particular interest is also the fact that tumor levels for $[^{67}\text{Ga}]\text{Ga-GAS3}$ remained unchanged between 1 and 4 h pi ($21.75 \pm 4.90\% \text{IA/g}$ at 1 h vs. $20.39 \pm 3.96\% \text{IA/g}$ at 4 h pi; $p > 0.05$). Furthermore, $[^{67}\text{Ga}]\text{Ga-GAS3}$, was cleared mainly via the kidneys, as opposed to $[^{67}\text{Ga}]\text{Ga-GAS2}$, which displayed high hepatobiliary excretion with unfavorably high radioactivity levels in the intestines at 1 and 4 h pi (>25%IA/g).

A similar trend was observed in the In-111 set of compounds, as far as the uptake in the HEK293-CCK_{2i4sv}R tumors at 4 h pi is concerned, and specifically: GAS3 ($19.83 \pm 1.35\%$ IA/g) > [¹¹¹In]In-GAS1 ($17.79 \pm 1.73\%$ IA/g; $p < 0.01$) >> [¹¹¹In]In-GAS2 ($8.91 \pm 1.59\%$ IA/g; $p < 0.0001$). Differences across radioligands were found more pronounced at 24 h pi, with [¹¹¹In]In-GAS3 ($11.53 \pm 1.93\%$ IA/g) > [¹¹¹In]In-GAS1 ($5.40 \pm 1.13\%$ IA/g; $p < 0.0001$) > [¹¹¹In]In-GAS2 ($2.78 \pm 0.43\%$ IA/g; $p < 0.0001$). Background clearance followed a similar trend to that of Ga-67 analogs. Thus, [¹¹¹In]In-GAS3 was predominantly cleared via the kidneys while [¹¹¹In]In-GAS2 displayed significant intestinal values at 4 h pi ($9.71 \pm 0.42\%$ IA/g). All three In-111 radioligands displayed notable renal uptake. In a pilot experiment, the kidney protection agent gelofusine was coinjected together with [¹¹¹In]In-GAS1/2 to detect a potential reduction of renal accumulation at both the 4 and 24 h pi intervals, reported previously for [^{99m}Tc]Tc-DGA1, [^{99m}Tc]Tc-DG2 and many other CCK₂R-targeting peptide radiotracers [38,44]. Surprisingly, gelofusine failed to reduce kidney uptake or any other pharmacokinetic parameter (Tables S6 and S7; Supplementary File).



(a)



(b)

Figure 4. Cont.

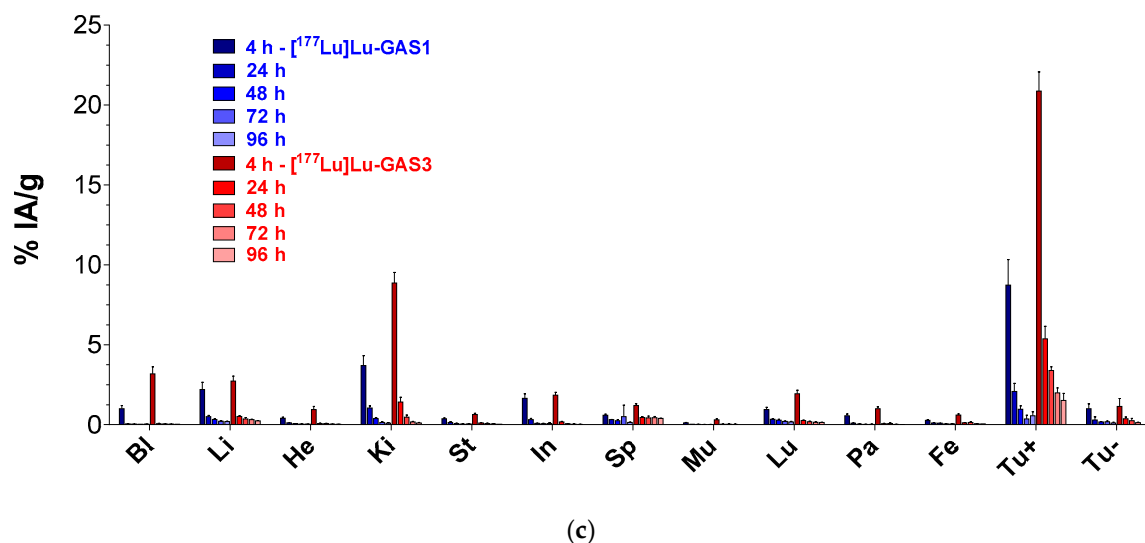


Figure 4. Comparative biodistribution data in SCID mice bearing twin HEK293-CCK_{2i4sv}R and wtHEK293 tumors in their flanks expressed as % IA/g tissue (mean \pm SD, $n = 4$) for (a) [⁶⁷Ga]Ga-GAS1/2/3, (b) [¹¹¹In]In-GAS1/2/3 and (c) [¹⁷⁷Lu]Lu-GAS1/3; Bl: blood, Li: liver, He: heart, Ki: kidneys, St: stomach, In: intestines, Sp: spleen, Mu: muscle, Lu: lungs, Pa: pancreas, Fe: femur, Tu+/-: HEK293-CCK_{2i4sv}R/wtHEK293 tumor.

Concordant with these findings, [¹⁷⁷Lu]Lu-GAS3 showed higher HEK293-CCK_{2i4sv}R tumor-targeting capacity compared with [¹⁷⁷Lu]Lu-GAS1 at all time intervals (Tables S9 and S10, respectively; Supplementary File). For example, [¹⁷⁷Lu]Lu-GAS3 > [¹⁷⁷Lu]Lu-GAS1 at 4 h pi ($20.88 \pm 1.20\%$ IA/g vs. $8.74 \pm 1.60\%$ IA/g; $p < 0.0001$) and at 24 h pi ($5.38 \pm 0.77\%$ IA/g vs. $2.09 \pm 0.50\%$ IA/g; $p < 0.0001$). However, tumor values significantly declined with time. On the other hand, they both showed rapid background clearance predominantly via the kidneys. The kidneys represented the organ with the highest activity uptake after the HEK293-CCK_{2i4sv}R tumors, but renal levels drastically dropped between 4 and 24 h pi (e.g., [¹⁷⁷Lu]Lu-GAS3 from $8.86 \pm 0.67\%$ IA/g to $1.42 \pm 0.29\%$ IA/g, respectively; $p < 0.0001$). It should be noted that for [¹⁷⁷Lu]Lu-GAS3 tumor to kidney favorably increased with time, e.g., from 2.4 at 4 h pi, to 3.8 at 24 h pi and up to 13.8 at 96 pi.

3.6. SPECT/CT with [¹¹¹In]In-GAS1/2/3

SPECT/CT imaging was performed for [¹¹¹In]In-GAS1/2/3 at 4 and 24 h pi in SCID mice bearing double HEK293-CCK_{2i4sv}R and wtHEK293 tumors in their flanks, and the results are summarized in Figure 5.

We observed that radioactivity was selectively taken up only by the HEK293-CCK_{2i4sv}R tumors, with the receptor-negative tumors remaining devoid of radioactivity for all compounds. Thus, concordant with biodistribution findings, these results reveal that tumor uptake of [¹¹¹In]In-GAS1/2/3 is a CCK_{2i4sv}R-mediated process. Furthermore, and in agreement with biodistribution results, [¹¹¹In]In-GAS3 displayed a superior pharmacokinetic profile.

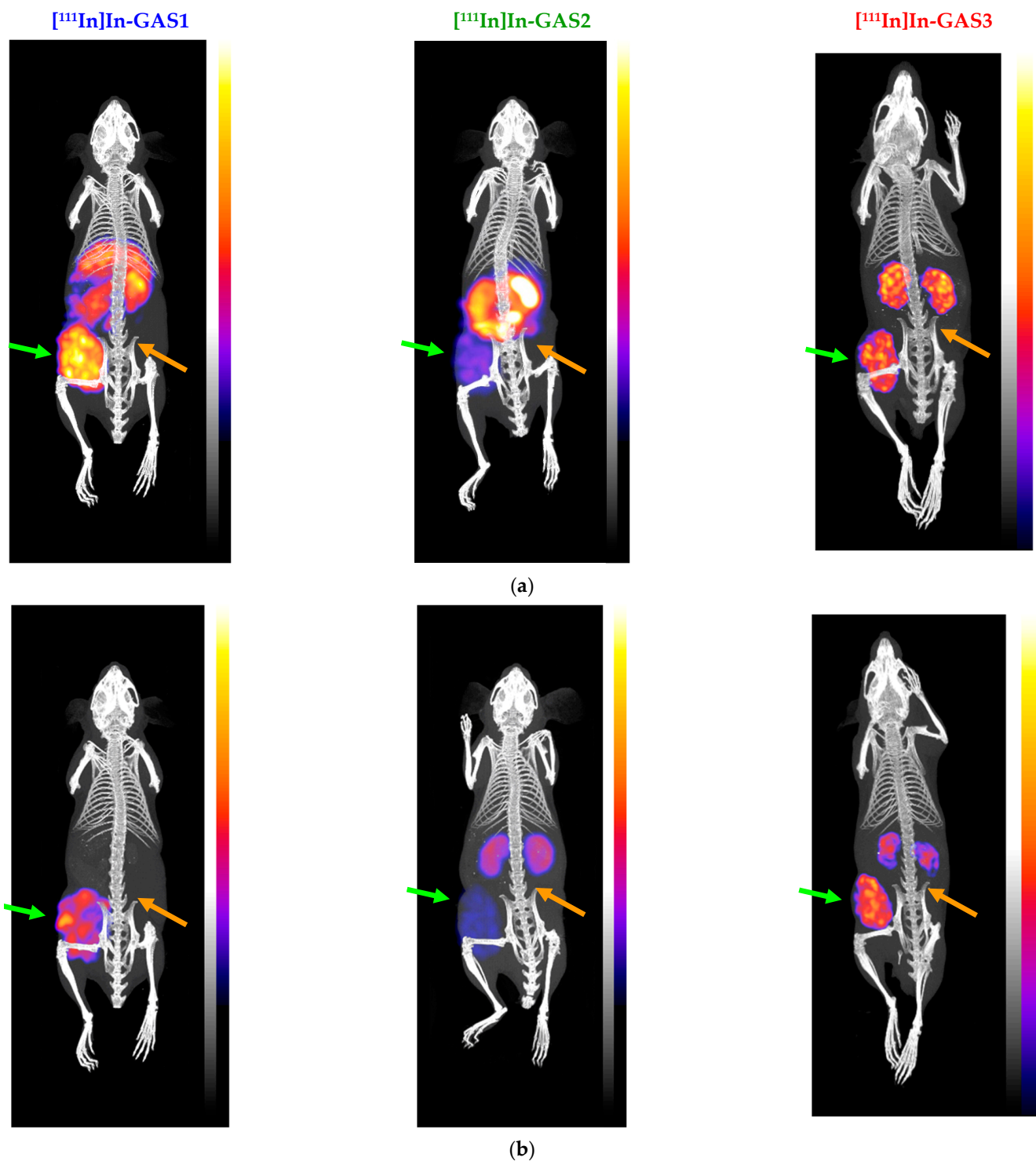


Figure 5. Static whole-body SPECT/CT images of SCID mice bearing twin HEK293-CCK_{2i4svR} and wtHEK293 tumors in their flanks at (a) at 4 h pi and (b) at 24 h pi of [¹¹¹In]In-GAS1/2/3; green arrows are pointing to HEK293-CCK_{2i4svR} xenografts, and orange arrows are indicating the wtHEK293 tumors. Intense uptake is observed in the CCK_{2i4svR} expressing tumors, but no uptake is evident in the tumors devoid of CCK_{2i4svR} expression. The color bars indicate the difference in accumulated activity (purple being the lowest and white being the highest level of accumulation).

4. Discussion

The development of gastrin-based anti-CCK₂R radioligands for the theranostic management of human tumors showed significant advances in recent years [8]. Despite this, a

few pending issues need to be tackled, for example, the side-effects elicited after CCK₂R activation by agonists injected to patients [25], or the inadvertent accumulation of radioactivity in tissues with high physiological CCK₂R-expression (e.g., the gastric mucosa) [11,22,23,42]. Therefore, a shift of paradigm toward radiolabeled CCK₂R-antagonists represents the next rational step to take in this venture, mimicking recent developments in the fields of somatostatin and bombesin [45,46]. Adopting this concept, we recently introduced [^{99m}Tc]Tc-DGA1, a radiotracer based on the nonpeptidic antagonist Z360 and suitable for visualizing CCK₂R-positive lesions with SPECT/CT [38]. Aiming toward Z360-radioligands with a broader theranostic profile, we now present GAS1/2/3. These Z360-conjugates were modified by three different macrocyclic chelators (DOTA, NODAGA, or DOTAGA) suitable for labeling with trivalent radiometals of clinical interest with each chelator tethered to the free carboxylate group of Z360 via the common linker H-DGlu-HN(PEG3)NH₂ (Figure 1). Hence, unlike DGA1, GAS1/2/3 lack the (DGlu)₄-chain implicated in the observed high renal uptake of [^{99m}Tc]Tc-DGA1 [38].

The CCK₂R-affinity of the three bioconjugates was found to be slightly inferior to the unmodified Z360. After the incorporation of gallium, indium or lutetium, more pronounced effects on CCK₂R-affinity could be observed across analogs, with the poorest affinity displayed by [^{nat}In]In-GAS2 (Table 1). Differences in the metal-chelate charge, polarity and overall configuration residing on distinct coordination chemistries of the aforementioned trivalent metals with the three chelators [47] seem to play a role in determining the CCK₂R-affinity of the resulting metal-tagged compounds. It should be noted that the GAS3 series displayed superior CCK₂R-affinities compared with the respective GAS1 and GAS2 counterparts.

Next, the GAS1/2/3 were studied for agonism–antagonism at the CCK₂R applying Ca²⁺ mobilization assays in HEK293-hCCK2R-YC3.60 cells. As expected, the two positive controls [Leu¹⁵]gastrin-17 and DG2, behaved as typical agonists, triggering Ca²⁺ mobilization at concentrations as low as 100 pM and 10 pM, respectively (Figure S1A,B; Supplementary File). At this concentration level, none of the GAS1/2/3 or the Z360 reference were able to induce a visible effect. Unexpectedly though, these analogs showed agonistic properties at concentrations >100 nM (Figures S1C and S2A–C; Supplementary File). Furthermore, during additional pilot experiments, they failed to inhibit Ca²⁺ mobilization induced by either [Leu¹⁵]gastrin-17 or DG2 agonists (results not shown), in contrast to DGA1 previously showing a pure antagonist profile at the CCK₂R [38]. The intriguing behavior of GAS1/2/3 and Z360 at the CCK₂R seems to be in line with an allosteric modulator profile, as reported for other benzodiazepine analogs [48,49], and requires dedicated studies to be fully explored. For nuclear medicine applications, the absence of CCK₂R-activation by GAS1/2/3 at concentrations <100 nM is significant and associated with sufficient safety for injection in humans. Notably, hitherto clinical studies, including those using therapeutic doses, involve the administration of CCK₂R-agonists well below 30 nmol/patient [22,23]. This would translate to a concentration of a maximum of 6 nM in a 75 kg patient (without taking into account the rapid blood clearance or the much higher distribution volume in the patient than the 5 Lt blood volume). This value is well below the threshold of CCK₂R-activation by any of GAS1/2/3.

Radiolabeled GAS1/2/3 displayed different receptor-specific uptake during incubation in HEK293-CCK₂4svR cells, but in general, cell uptake followed binding affinity patterns. For example, the most affine [¹¹¹In]In-GAS3 displayed the highest cell-uptake values at 1 h and [¹¹¹In]In-GAS2 the poorest (Figure 3 and Table 1). Another interesting feature in the cell uptake of these radioligands is the varying distribution of activity between the internalized and membrane-bound fragments across analogs. Thus, the internalized to membrane-bound ratio was as high as 4.1 for [¹¹¹In]In-GAS3 and 2.8 for [¹⁷⁷Lu]Lu-GAS3, but dropped to considerably lower values for [⁶⁷Ga]Ga-GAS1 (0.7) and [⁶⁷Ga]Ga-GAS2 (0.9). The observed cell-uptake and internalization pattern is not consistent with a typical receptor antagonist and evokes the unlikely behavior of GAS1/2/3 during the Ca²⁺-mobilization assays. Previous reports showed benzodiazepine analogs behaving

as antagonists during Ca^{2+} mobilization functional assays while at the same time inducing partial or full CCK_2R -internalization in different assays [50,51].

Following injection in healthy mice, the radioligands displayed high metabolic stability in peripheral mice blood at 5 min pi and, with the exception of ^{111}In In-GAS2, were found excreted without further degradation in the urine at 30 min pi (Figure 3 and Table 2). This finding represents an advantageous trait of the new nonpeptidic Z360-radioligands vs. their peptidic counterparts based on gastrin. The latter, in particular the *des*-(Glu)₅ analogs, were shown to undergo fast enzymatic degradation after entering the bloodstream by NEP and possibly also ACE [11–15,43,52,53]. It should be noted that radiopeptide metabolic stability improvements could be achieved either via in situ NEP and ACE inhibition strategies or after extensive structural interventions of peptide motifs [8].

The tumor-targeting capabilities and overall biodistribution patterns of GAS1/2/3-radioligands were studied in immunosuppressed mice bearing a twin xenograft model, namely subcutaneous tumors from either HEK293- $\text{CCK}_{2i4sv}\text{R}$ or wtHEK293 cells (Figure 4, Tables S3–S9 in Supplementary File). We observed that biodistribution profiles depend on radiometal-chelate, with the targeting of HEK293- $\text{CCK}_{2i4sv}\text{R}$ tumors being superior in the case of Ga-67 and In-111 labeled GAS1 and GAS3, compared with GAS2. Furthermore, ^{67}Ga Ga-GAS2 displayed significantly higher intestinal accumulation vs. both ^{67}Ga Ga-GAS1/3, while ^{111}In In-GAS2 showed unfavorably higher uptake in both the kidneys and intestines vs. the ^{111}In In-GAS1/3 counterparts. Overall, the NODAGA-Z360-based ^{67}Ga Ga/ ^{111}In In-radioligands showed the least favorable tumor-to-background profile vs. the DOTA/DOTAGA-modified radioligands. On the contrary, GAS1 and GAS3 displayed high tumor targeting and lower background levels after labeling with either Ga-67, In-111, or Lu-177. Amongst the two, ^{67}Ga Ga/ ^{111}In In/ ^{177}Lu Lu-GAS3 displayed a superior profile in mice than the corresponding ^{67}Ga Ga/ ^{111}In In/ ^{177}Lu Lu-GAS1. In the case of the ^{111}In In-GAS1/2/3 radioligands, these results were nicely illustrated by SPECT/CT (Figure 5).

The present study revealed a few promising features with regards to the application of ^{177}Lu Lu-GAS1/3 as anti- CCK_2R tumor therapeutics. Clearly, washout from the experimental tumors from 4 to 24 h pi is concerning; this uptake gradually declined further at the later time points. However, background activity declined notably faster, resulting in positive tumor-to-background ratios over time, a finding in support of future therapy prospects. Such prospects should be further validated by future systematic therapy studies in mice, aiming to address a number of questions still open at this preliminary stage. On the one hand, Z360-conjugate/radionuclide doses and administration schemes need to be explored and optimized to maximize therapeutic efficacy. Then, potential damage to healthy tissues needs to be carefully investigated by dedicated immune-histological assays and reliable dosimetry calculations to be performed before setting toxicity limits. In this respect, the lack of CCK_2R -activation at concentrations <100 nM, along with the low radioactivity levels of the new analogs in the stomach, represent clear advantages. Carefully conducted therapy studies at the preclinical level will provide crucial information on the therapeutic value of either ^{177}Lu Lu-GAS1/3 per se, but may also hint at the need for further modifications on the radioligand structure (with focus on the linker) or the application of alternative to Lu-177 particle emitters with better suited nuclear properties [47].

Compared with the previously reported biodistribution data of ^{99m}Tc Tc-DGA1 in the same animal model, we observed a much higher tumor to kidney ratio for all ^{67}Ga Ga/ ^{111}In In/ ^{177}Lu Lu-GAS1/3 radioligands and an overall improvement of the pharmacokinetic profiles [38]. Of particular interest is the fact that mice treatment with gelofusine, previously shown to significantly reduce the kidney accumulation of ^{99m}Tc Tc-DGA1 (from $96.09 \pm 12.01\%$ IA/g to $33.61 \pm 2.67\%$ IA/g at 4 h pi; $p < 0.0001$), had no effect on the renal uptake of ^{111}In In-GAS1 ($6.06 \pm 1.24\%$ IA/g vs. gelofusine $5.93 \pm 0.56\%$ IA/g at 4 h pi; $p > 0.05$) or ^{111}In In-GAS2 ($29.55 \pm 3.64\%$ IA/g vs. gelofusine $21.90 \pm 3.97\%$ IA/g at 4 h pi; $p > 0.05$), correlating the effect of the kidney protection agent with the (DGlu)₄-chain in ^{99m}Tc Tc-DGA1.

Another quite attractive feature of radiolabeled GAS1/2/3, in line with a radioantagonist profile, is the low uptake in the CCK₂R-rich stomach. As mentioned above, this feature becomes particularly important for therapeutic purposes due to dosimetric restrictions [8]. Interestingly, the injection of higher peptide amounts was shown to reduce stomach uptake in preclinical models, but it is associated with more biosafety risks in the case of CCK₂R-agonists [54].

Although a hitherto restricted number of other Z360 analogs carrying the DOTA-chelator through different linkers was developed [55], animal data was only reported for [¹¹¹In]In-IP-001, but in a mice model different to ours [56]. Thus, an A549 non-small cell lung cancer xenograft was implanted in the shoulder of homozygous female BALB/c nude mice. Specific but low uptake could be established in the tumor on SPECT/CT 4 and 24 h after injection of the radioligand combined with an unfavorably high radioactivity background compared with the scans of Figure 5 and the biodistribution results of this study, especially for the GAS3 (and to a lesser extent the GAS1) radioligand series.

5. Conclusions

In the present work, three new bioconjugates of the benzodiazepine-like CCK₂R-antagonist Z360 were introduced, each coupled with one of the DOTA, NODAGA and DOTAGA chelators via a common linker, and amenable for stable labeling with clinically interesting trivalent radiometals, such as Ga-67 (surrogate for the PET radiometal Ga-68), In-111 and Lu-177 included in our study, and combining several attractive features. Firstly, they displayed receptor affinities dependent upon the metal-chelate, with a few members showing single-digit nM receptor affinities. During functional assays at the cellular level, none of the bioconjugates activated the CCK₂R at clinically relevant concentrations, thereby providing a dose-safety window for future human applications. The nonpeptidic Z360-radioconjugates tested herein displayed excellent *in vivo* stability in mice, as opposed to their gastrin-based peptidic counterparts undergoing fast *in vivo* degradation by NEP and sometimes also ACE. As a result, the new Z360 radioligands were able to specifically target CCK₂R-positive xenografts in mice but not CCK₂R-negative control tumors and displayed a low uptake in the CCK₂R-rich stomach, as consistent with receptor radioantagonists. The background clearance was fast, especially for the GAS3 (DOTAGA-carrying member) and to a lesser extent for the GAS1 (DOTA-carrying member) radioligand series. These excellent qualities confirm previous experience with somatostatin and bombesin antagonist radioligands, paving the way for the first therapy and clinical translation studies of CCK₂R-antagonists in MTC and other tumor patients.

Supplementary Materials: The following supporting information can be downloaded at: <https://www.mdpi.com/article/10.3390/pharmaceutics14030666/s1>, Analytical data for GAS1/2/3, Table S1: Analytical data for GAS1/2/3; Labeling of GAS-compounds with Ga-67, In-111 and Lu-177; Preparation of [^{nat}Ga]Ga-GAS1/2/3, [^{nat}In]In-GAS1/2/3 and [^{nat}Lu]Lu-GAS1/3; Table S2. Analytical HPLC data for (radio)metal-tagged GAS1/2/3; Ca²⁺ mobilization assays; Figure S1: Ca²⁺ mobilization assays for [Leu15]gastrin-17, DG2, and Z360 references in HEK293 cells contingent on the expression of CCK₂R; Figure S2: Ca²⁺ mobilization assays for GAS1/2/3 in HEK293 cells contingent on the expression of CCK₂R; Tables S3–S10: Biodistribution results for GAS1/2/3 labeled with Ga-67, In-111 and Lu-177 in SCID mice bearing double HEK293-CCK_{2i4sv}R/wtHEK293 tumors. References [57,58] are cited in the supplementary materials.

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Supplementary Materials: Nonpeptidic Z360-Analogs Tagged with Trivalent Radiometals as Anti-CCK₂R Cancer Theranostic Agents: A Preclinical Study

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Analytical data for GAS1/2/3

Analytical data from the synthesis of GAS1/2/3, comprising results from MALDI-TOF mass spectroscopy and from HPLC analysis in two distinct HPLC systems, were compiled in Table S1.

Table S1. Analytical Data for GAS1/2/3.

compound	structure	% purity ^a	MW ^b calcd	HPLC <i>t_R</i> (min) UV trace	
			MW ^b found, m/z	system 1 ^c	system 2 ^d
GAS1	DOTA-DGlu-HN(PEG) ₃ NH-Z360	≥ 95	1210.4/1210.9, [M + H] ⁺	27.0	17.0
GAS2	NODAGA-DGlu-HN(PEG) ₃ NH-Z360	≥ 95	1181.3/1181.9, [M + H] ⁺	36.1	17.6
GAS3	DOTAGA-DGlu-HN(PEG) ₃ NH-Z360	≥ 95	1282.4//1283.0, [M + H] ⁺	29.0	24.5 ^e

^aPurity was determined by HPLC systems 1 and 2. ^bAverage mass. ^csystem 1: RP-HPLC with UV detection at 220 nm; a Symmetry Shield RP18 column (5 μm, 3.9 mm × 150 mm) was eluted at a flow rate of 1 mL/min with linear gradient: 100%A/0% B to 70%A/30% B in 5 min and then 70%A/30% B to 55%A/45% B in 60 min, with A = 0.1% TFA in H₂O (v/v) and B = MeCN. ^dsystem 2: A Nucleosil-100 C18 column (5 μm, 150 mm × 4 mm) was eluted at a flow rate of 1 mL/min with a linear gradient: 0.1 % TFA in MeCN (10% to 90% in 15 min) and 0.1 % aqueous TFA as complementary phase; runs were monitored by UV detection at 215 nm; ^esystem 2 modified as: 10% to 90% in 50 min.

Labeling of GAS-compounds with Ga-67, In-111 and Lu-177

Labeling with Ga-67. To an Eppendorf Protein LoBind[®] centrifuge tube (capacity: 1.5 mL) the following items were successively added: EtOH (40 μL), [⁶⁷Ga]GaCl₃ (27 μL; 4 – 5.5 GBq/mL in dilute HCl), a freshly prepared solution of sodium para-aminobenzoic acid (Na-PABA, 10 μL, 0.5 M) in bi-distilled water, the corresponding GAS1/2/3 precursor stock solution (10 μL, 20 μg, ≈10 nmol) and sodium acetate solution (10 μL, pH 8, 1 M). The mixture was incubated for 30 min at 80 °C (for GAS1 and GAS3) or 40 °C (for GAS2). For quality control a 2 μL aliquot of the labeling solution was withdrawn and quenched with 28 μL of an acetate buffered solution of disodium ethylenediaminetetraacetic acid (Na₂-EDTA, 1 mM, pH 4.0). The [⁶⁷Ga]Ga-GAS1/2/3 radioligands were produced in radiochemical purities generally exceeding 95% at apparent molecular activities of 10 – 15 MBq [⁶⁷Ga]Ga/nmol GAS conjugate.

Labeling with In-111. Into an Eppendorf Protein LoBind[®] centrifuge tube (capacity: 1.5 mL) the following items were successively pipetted: EtOH (100 μL), [¹¹¹In]InCl₃ (150 μL, 400 – 600 MBq/mL in 0.05 M HCl) and a freshly prepared Na-PABA solution (25 μL, 0.5 M). After adding the corresponding GAS conjugate stock solution (25 μL, 50 μg, ≈25 nmol) and sodium acetate buffer (15 μL, pH 4.0, 1 M) the labeling reaction mixture was incubated as follows: GAS1 – at 50 °C for 70 min; GAS2 – at 50 °C for 50 min; GAS3 – at 80 °C for 30 min. Quality control was performed after adding

octadentate DTPA as a scavenger of unbound In-111. The [¹¹¹In]In-GAS1/2/3 radioligands were obtained in radiochemical purities exceeding 95 % at apparent molecular activities of 2.4 – 3.6 MBq [¹¹¹In]In/nmol GAS conjugate.

Labeling with Lu-177. For labeling of GAS1/3 with Lu-177, the following reagent solutions were pipetted into an Eppendorf Protein LoBind® centrifuge tube (capacity: 1.5 mL): EtOH (120 µL), [¹⁷⁷Lu]LuCl₃ (150 µL, 3.7 GBq/mL in 0.04 M HCl, A_s > 370 GBq/mg Lu), sodium acetate buffer (30 µL pH 4.6, 1.0 M), a freshly prepared Na-PABA solution (30 µL, 0.5 M) and each of the GAS1/3 stock solution (35 µL, 70 µg, ≈35 nmol). The mixture was heated at 80 °C for 20 min. Quality control was performed after adding octadentate DTPA as a scavenger of unbound Lu-177. [¹⁷⁷Lu]Lu-GAS1/3 were produced in high radiochemical purities (>95 %) at apparent molar activities of approximately 16 MBq [¹⁷⁷Lu]Lu/nmol ligand at the end of synthesis.

Results from radio-HPLC analysis applying system 1 were compiled in Table S2. In view of the high purity formation of radioligands, no further purification was required for subsequent biological testing. For all further *in vitro* and *in vivo* studies the labeling solution was diluted to the desired activity concentration using a 5 mM solution of Na-PABA dissolved in phosphate buffered saline. The integrity of radiotracers was tested before and after the conclusion of all biological experiments.

Table S2. Analytical HPLC Data for (Radio)Metal-Tagged GAS1/2/3 (*t_R* (min) gamma/UV trace).

compound	-	[⁶⁷ Ga]Ga/[^{nat} Ga]Ga	[¹¹¹ In]In/[^{nat} In]In	[¹⁷⁷ Lu]Lu/[^{nat} Lu]Lu
GAS1	27.0	30.0	31.7	33.1
GAS2	36.1	36.3	34.8	-
GAS3	29.0	30.3	36.2	39.3

Purity as determined by HPLC exceeded 97% in all cases. system 1 was applied: RP-HPLC with twin UV detection at 220 nm and gamma detection; a Symmetry Shield RP18 column (5 µm, 3.9 mm × 150 mm) was eluted at a flow rate of 1 mL/min with linear gradient: 100%A/0% B to 70%A/30% B in 5 min and then 70%A/30% B to 55%A/45% B in 60 min, with A = 0.1% TFA in H₂O (v/v) and B = MeCN.

Preparation of [^{nat}Ga]Ga-GAS1/2/3, [^{nat}In]In-GAS1/2/3 and [^{nat}Lu]Lu-GAS1/3

To an Eppendorf Protein LoBind® centrifuge tube containing GAS1/2/3 precursor stock solution (60 µL, 120 µg, ≈60 nmol), a stock solution (60 µL, 2 mM, 120 nmol) of the respective nitrate salt dissolved in 1 M sodium acetate buffer of pH 4.0 (for gallium), 4.6 (for indium) and 5.0 (for lutetium) was added. The mixture was heated at 75 °C for 1 h. Complete metal incorporation by GAS1/2/3 was revealed by RP-HPLC analysis of the reaction mixture using HPLC system 1, allowing for base line separation of metal-tagged from metal-free GAS1/2/3. Retention times (UV trace, *t_R* in min) of all GAS species were included in Table S2. The metal-tagged GAS solutions were used as such in *in vitro* competition binding assays assuming a 0.5 mM CCK₂R-ligand concentration.

Ca²⁺ Mobilization Assays

Methods for FRET-based detection of Ca²⁺ in HEK293 cells were described previously [1]. The clone #21 HEK293 cell line (HEK293-hCCK2R/YC3.60) stably co-expressing the human CCK₂R and the Ca²⁺ biosensor YC3.60 was generated according to published methods [2]. The clone #5 HEK293 cell line (HEK293-YC3.60) that expressed only YC3.60 served as a negative control and was generated in a similar matter. The live-cell Ca²⁺ assay was performed in a 96-well format using suspensions of HEK293-

hCCK₂R/YC3.60 cells to which test compounds were administered by automated injection using a Flexstation 3 microspectrofluorimeter (Molecular Devices Corp.). The human CCK₂R (NM_176875) in pCMV6-Entry vector contained the receptor's complete open reading frame with a Myc-DDK epitope tag (OriGene Technologies; Cat. No. R200676). Test compounds included human [Leu¹⁵]-gastrin-17 (BACHEM Cat. No. 4011661), Z360 (MedChem Express Cat. No. HY-17617), Demogastin-2 (DG2) and GAS1/2/3 (PiChem Forschungs- und Entwicklungs GmbH, Raaba-Grambach, Austria). ADP (Sigma-Aldrich) served as a positive control test compound for intracellular Ca²⁺ mobilization.

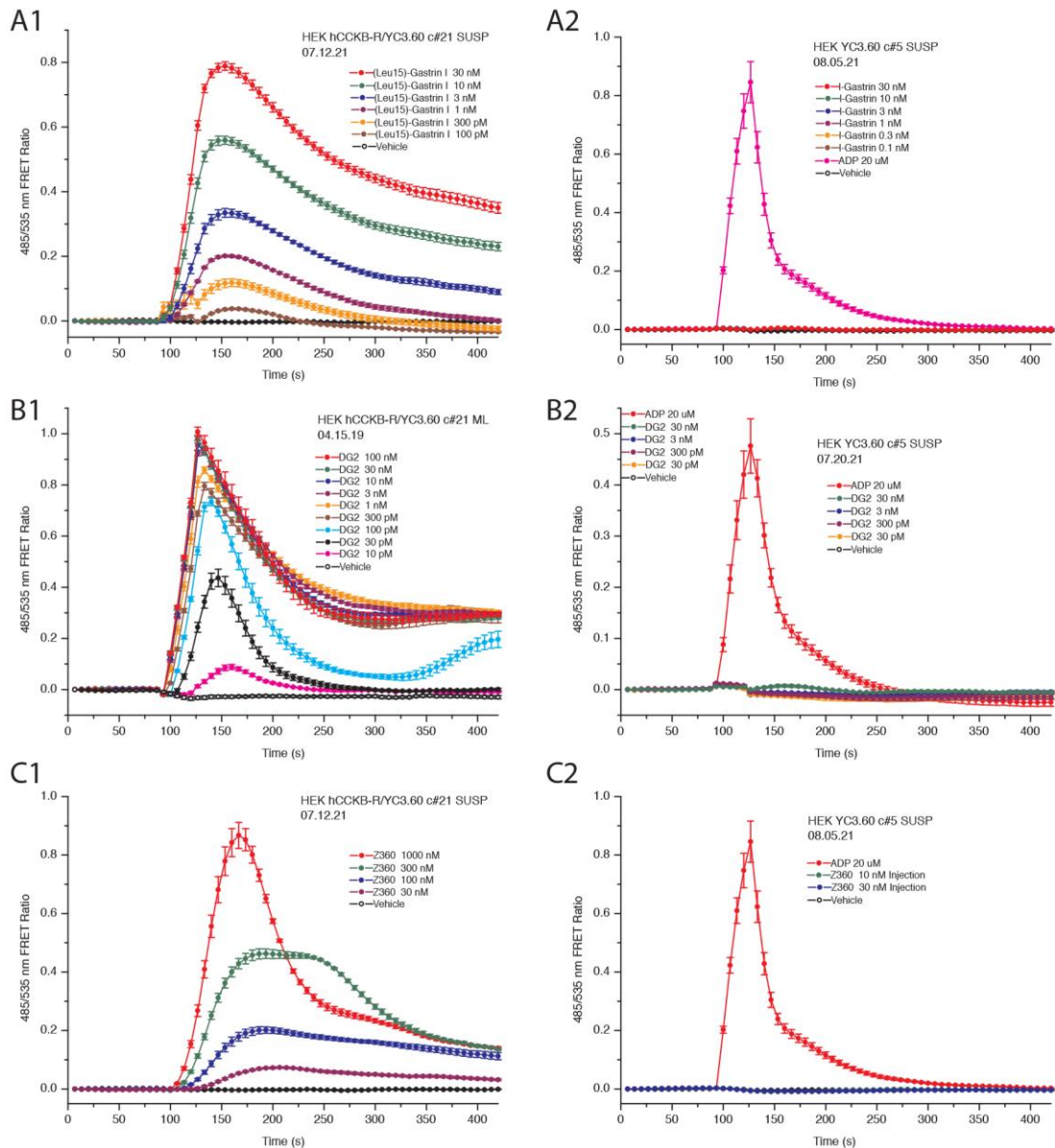


Figure S1. Illustrated are concentration-response relationships for test compounds obtained in YC3.60 FRET-based Ca²⁺ mobilization assays using clone #21 HEK293-hCCK₂R-YC3.60 cells (A1, B1, C1) or clone #5 HEK293-YC3.60 cells that do not express CCK₂R (A2, B2, C2). Panels A1 and A2 demonstrate that a Ca²⁺ mobilizing action of [Leu¹⁵]gastrin-17 was observed using clone #21 but not clone #5 cells. Note that although clone #5 cells failed to respond to [Leu¹⁵]gastrin-17, they instead responded to ADP that stimulated endogenous purinergic receptors expressed on HEK293 cells. Panels B1 and B2 illustrate findings obtained when testing DG2, whereas

panels C1 and C2 illustrate findings obtained when testing Z360. Collectively, these findings demonstrate that the Ca^{2+} mobilizing actions of [Leu¹⁵]gastrin-17, DG2, and Z360 in HEK293 cells are contingent on expression of CCK₂R.

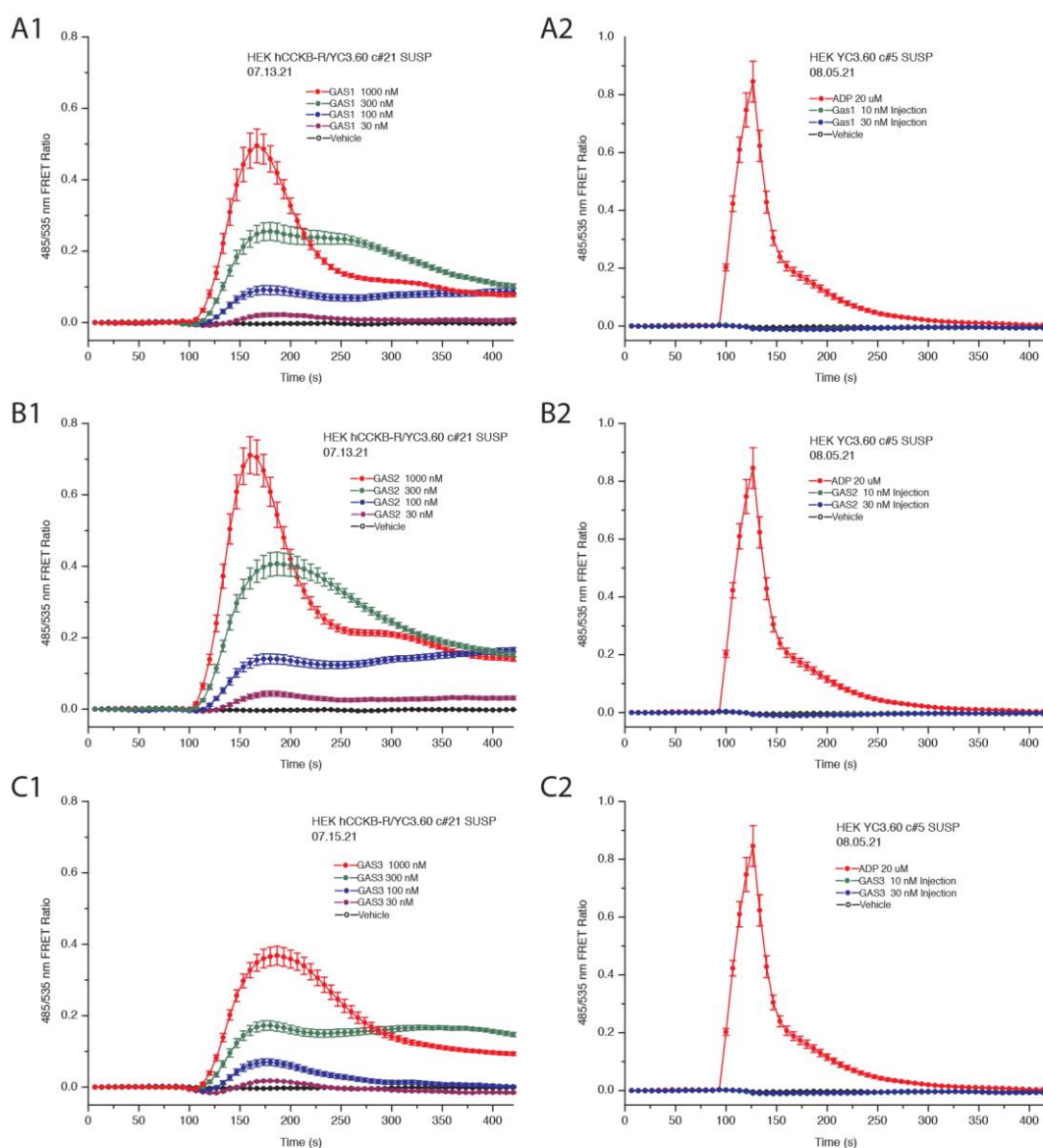


Figure S2. Illustrated are concentration-response relationships for test compounds obtained in assays using clone #21 HEK293-hCCK₂R-YC3.60 cells (A1, B1, C1) or clone #5 HEK293-YC3.60 cells (A2, B2, C2). Panels A1 and A2 demonstrate a Ca^{2+} mobilizing action of GAS1 using clone #21 cells but not clone #5 cells. Thus, the agonist action of GAS1 was contingent on expression of CCK₂R. Similar findings were obtained when testing GAS2 (B1, B2) or GAS3 (C1, C2).

Biodistribution in Mice Bearing Twin HEK293-CCK₂svR and wtHEK293 Xenografts

Biodistribution results for GAS1/2/3 labeled with Ga-67, In-111 and Lu-177 in SCID mice bearing double HEK293-CCK₂svR / wtHEK293 tumors, expressed as %IA/g and representing average values \pm SD, n= 4, per animal group, were included in the following Tables S3-S10.

Table S3. Biodistribution of [⁶⁷Ga]Ga-GAS1 in male SCID mice bearing twin HEK293-CCK2_{4sv}R and wtHEK293 tumors at 1, 4 and 24 h pi, expressed as %IA/g and representing mean values ± SD, n=4.

Organs	%IA/g ± SD, n=4		
	1 h	4 h	24 h
Blood	6.53 ± 1.33	1.25 ± 0.27	0.08 ± 0.01
Liver	3.39 ± 0.35	2.49 ± 0.31	0.75 ± 0.08
Heart	2.11 ± 0.86	0.47 ± 0.09	0.13 ± 0.01
Kidneys	5.78 ± 0.48	3.59 ± 0.60	0.88 ± 0.17
Stomach	1.09 ± 0.22	0.37 ± 0.08	0.18 ± 0.02
Intestines	2.33 ± 0.44	2.84 ± 0.38	0.32 ± 0.13
Spleen	1.63 ± 0.16	0.78 ± 0.02	0.51 ± 0.12
Muscle	0.61 ± 0.07	0.19 ± 0.07	0.05 ± 0.02
Lungs	3.63 ± 0.69	1.19 ± 0.21	0.31 ± 0.16
Femur	0.97 ± 0.12	0.29 ± 0.16	0.18 ± 0.04
Pancreas	2.26 ± 0.18	0.51 ± 0.09	0.22 ± 0.01
CCK _{24sv} R(+) Tumor	12.62 ± 1.76	9.55 ± 1.344.59	2.47 ± 0.41
CCK _{24sv} R(-) Tumor	1.62 ± 0.12	0.75 ± 0.19	0.00 ± 0.00

Table S4. Biodistribution of [⁶⁷Ga]Ga-GAS2 in male SCID mice bearing twin HEK293-CCK2_{4sv}R and wtHEK293 tumors at 1, 4 and 24 h pi, expressed as %IA/g and representing mean values ± SD, n=4.

Organs	%IA/g ± SD, n=4		
	1 h	4 h	24 h
Blood	0.86 ± 0.08	0.19 ± 0.06	0.05 ± 0.00
Liver	5.51 ± 2.03	2.64 ± 0.85	0.78 ± 0.17
Heart	0.27 ± 0.03	0.10 ± 0.03	0.04 ± 0.01
Kidneys	1.48 ± 0.18	0.39 ± 0.11	0.06 ± 0.01
Stomach	0.32 ± 0.14	0.14 ± 0.06	0.06 ± 0.02
Intestines	25.26 ± 2.90	26.38 ± 11.89	0.31 ± 0.32
Spleen	0.52 ± 0.06	0.55 ± 0.15	0.35 ± 0.02
Muscle	0.12 ± 0.02	0.04 ± 0.01	0.02 ± 0.00
Lungs	0.80 ± 0.09	0.34 ± 0.01	0.15 ± 0.02
Femur	0.24 ± 0.03	0.13 ± 0.03	0.10 ± 0.03
Pancreas	0.37 ± 0.05	0.08 ± 0.05	0.03 ± 0.00
CCK _{24sv} R(+) Tumor	4.06 ± 0.07	1.53 ± 0.26	0.24 ± 0.03
CCK _{24sv} R(-) Tumor	0.48 ± 0.03	0.19 ± 0.08	0.08 ± 0.00

Table S5. Biodistribution of [⁶⁷Ga]Ga-GAS3 in male SCID mice bearing twin HEK293-CCK2_{2i4sv}R and wtHEK293 tumors at 1, 4 and 24 h pi, expressed as %IA/g and representing mean values ± SD, n=4.

Organs	%IA/g ± SD, n=4		
	1 h	4 h	24 h
Blood	8.61 ± 1.17	2.23 ± 0.39	0.14 ± 0.02
Liver	5.62 ± 0.40	4.97 ± 0.74	2.48 ± 0.34
Heart	2.23 ± 0.38	0.76 ± 0.07	0.17 ± 0.02
Kidneys	9.09 ± 1.51	7.86 ± 0.48	1.98 ± 0.45
Stomach	1.51 ± 0.13	0.61 ± 0.12	0.19 ± 0.03
Intestines	1.53 ± 0.19	1.59 ± 0.19	0.39 ± 0.06
Spleen	1.96 ± 0.13	1.60 ± 0.39	1.67 ± 0.37
Muscle	0.82 ± 0.14	0.25 ± 0.05	0.06 ± 0.00
Lungs	4.32 ± 0.58	1.51 ± 0.22	2.43 ± 1.04
Femur	1.60 ± 0.31	0.78 ± 0.10	0.52 ± 0.07
Pancreas	2.51 ± 0.22	0.77 ± 0.11	0.17 ± 0.03
CCK _{2i4sv} R(+) Tumor	21.75 ± 4.90	20.39 ± 3.96	5.37 ± 1.00
CCK _{2i4sv} R(-) Tumor	2.52 ± 0.28	1.15 ± 0.18	0.61 ± 0.08

Table S6. Biodistribution of [¹¹¹In]In-GAS1 in male SCID mice bearing twin HEK293-CCK2_{2i4sv}R and wtHEK293 tumors at 4 and 24 h pi (control and Gelofusine-treated mice), expressed as %IA/g and representing mean values ± SD, n=4.

Organs	%IA/g ± SD, n=4			
	4 h	4 h + Gelo	24 h	24 h + Gelo
Blood	3.51 ± 0.56	2.75 ± 0.48	0.08 ± 0.01	0.07 ± 0.01
Liver	7.69 ± 1.00	6.34 ± 0.86	2.27 ± 0.30	1.30 ± 0.15
Heart	1.10 ± 0.19	0.92 ± 0.16	0.15 ± 0.03	0.15 ± 0.02
Kidneys	6.06 ± 1.24	5.93 ± 0.56	1.84 ± 0.38	1.69 ± 0.13
Stomach	0.98 ± 0.21	0.77 ± 0.14	0.21 ± 0.05	0.25 ± 0.04
Intestines	3.37 ± 0.89	1.87 ± 0.08	0.59 ± 0.11	0.49 ± 0.13
Spleen	1.56 ± 0.22	1.13 ± 0.28	0.64 ± 0.12	0.48 ± 0.08
Muscle	0.26 ± 0.04	0.28 ± 0.06	0.06 ± 0.00	0.06 ± 0.01
Lungs	2.23 ± 0.39	1.86 ± 0.22	0.36 ± 0.05	0.39 ± 0.03
Femur	0.77 ± 0.13	0.65 ± 0.15	0.16 ± 0.03	0.16 ± 0.01
Pancreas	1.49 ± 0.19	1.47 ± 0.26	0.18 ± 0.03	0.14 ± 0.01
CCK _{2i4sv} R(+) Tumor	17.79 ± 1.73	19.17 ± 2.06	5.40 ± 1.13	4.51 ± 0.55
CCK _{2i4sv} R(-) Tumor	0.73 ± 0.05	0.86 ± 0.15	0.27 ± 0.10	0.23 ± 0.04

Table S7. Biodistribution of [¹¹¹In]In-GAS2 in male SCID mice bearing twin HEK293-CCK_{2i4sv}R and wtHEK293 tumors at 4 and 24 h pi (control and Gelofusine-treated mice), expressed as %IA/g and representing mean values ± SD, n=4.

Organs	%IA/g ± SD, n=4			
	4 h	4 h + Gelo	24 h	24 h + Gelo
Blood	6.40 ± 0.75	6.29 ± 1.08	0.13 ± 0.02	0.15 ± 0.02
Liver	5.42 ± 0.89	4.56 ± 0.49	0.96 ± 0.22	0.95 ± 0.12
Heart	1.61 ± 0.23	1.42 ± 0.21	0.22 ± 0.01	0.22 ± 0.03
Kidneys	29.55 ± 3.64	21.90 ± 3.97	10.16 ± 2.30	11.51 ± 2.64
Stomach	1.31 ± 0.21	0.94 ± 0.15	0.36 ± 0.14	0.35 ± 0.06
Intestines	9.71 ± 0.42	9.48 ± 0.99	0.68 ± 0.15	0.75 ± 0.06
Spleen	1.49 ± 0.14	1.32 ± 0.20	0.51 ± 0.09	0.56 ± 0.17
Muscle	0.56 ± 0.08	0.47 ± 0.04	0.08 ± 0.01	0.08 ± 0.01
Lungs	3.27 ± 0.46	2.72 ± 0.30	0.60 ± 0.37	0.44 ± 0.07
Femur	1.15 ± 0.10	0.99 ± 0.17	0.24 ± 0.04	0.25 ± 0.03
Pancreas	2.99 ± 0.96	3.00 ± 0.58	0.29 ± 0.03	0.30 ± 0.05
CCK _{2i4sv} R(+) Tumor	8.91 ± 1.59	7.67 ± 0.61	2.78 ± 0.43	2.92 ± 0.37
CCK _{2i4sv} R(-) Tumor	1.82 ± 0.25	1.72 ± 0.30	0.41 ± 0.10	0.45 ± 0.09

Table S8. Biodistribution of [¹¹¹In]In-GAS3 in male SCID mice bearing twin HEK293-CCK_{2i4sv}R and wtHEK293 tumors at 4 and 24 h pi, expressed as %IA/g and representing mean values ± SD, n=4.

Organs	%IA/g ± SD, n=4	
	4 h	24 h
Blood	2.42 ± 0.94	0.08 ± 0.01
Liver	2.78 ± 0.27	1.07 ± 0.26
Heart	0.81 ± 0.28	0.16 ± 0.02
Kidneys	9.32 ± 0.68	4.15 ± 1.06
Stomach	0.91 ± 0.39	0.35 ± 0.21
Intestines	1.66 ± 0.33	0.48 ± 0.14
Spleen	0.69 ± 0.12	0.43 ± 0.09
Muscle	0.31 ± 0.13	0.08 ± 0.01
Lungs	1.36 ± 0.37	0.34 ± 0.07
Femur	0.60 ± 0.14	0.27 ± 0.09
Pancreas	0.86 ± 0.23	0.17 ± 0.04
CCK _{2i4sv} R(+) Tumor	19.83 ± 1.35	11.53 ± 1.93
CCK _{2i4sv} R(-) Tumor	0.85 ± 0.37	0.40 ± 0.21

Table S9. Biodistribution of [¹⁷⁷Lu]Lu-GAS1 in male SCID mice bearing twin HEK293-CCK2i4svR and wtHEK293 tumors at 4, 24, 48, 72 and 96 h pi, expressed as %IA/g and representing mean values ± SD, n=4.

Organs	%IA/g ± SD, n=4				
	4 h	24 h	48 h	72 h	96 h
Blood	0.99 ± 0.20	0.06 ± 0.00	0.04 ± 0.01	0.04 ± 0.00	0.04 ± 0.00
Liver	2.21 ± 0.44	0.51 ± 0.07	0.32 ± 0.04	0.23 ± 0.02	0.19 ± 0.02
Heart	0.41 ± 0.07	0.12 ± 0.00	0.07 ± 0.01	0.05 ± 0.01	0.04 ± 0.00
Kidneys	3.70 ± 0.62	1.05 ± 0.12	0.38 ± 0.07	0.14 ± 0.04	0.10 ± 0.04
Stomach	0.37 ± 0.05	0.14 ± 0.03	0.08 ± 0.01	0.05 ± 0.01	0.05 ± 0.01
Intestines	1.65 ± 0.28	0.32 ± 0.07	0.08 ± 0.01	0.06 ± 0.02	0.08 ± 0.05
Spleen	0.59 ± 0.07	0.32 ± 0.01	0.26 ± 0.06	0.51 ± 0.71	0.14 ± 0.02
Muscle	0.12 ± 0.02	0.04 ± 0.00	0.03 ± 0.00	0.02 ± 0.00	0.02 ± 0.01
Lungs	0.94 ± 0.15	0.34 ± 0.03	0.29 ± 0.04	0.21 ± 0.03	0.17 ± 0.02
Femur	0.28 ± 0.03	0.11 ± 0.01	0.09 ± 0.01	0.06 ± 0.00	0.05 ± 0.01
Pancreas	0.56 ± 0.11	0.10 ± 0.01	0.05 ± 0.01	0.03 ± 0.00	0.03 ± 0.01
CCK2i4svR(+) Tumor	8.74 ± 1.60	2.09 ± 0.50	0.97 ± 0.20	0.35 ± 0.23	0.56 ± 0.25
CCK2i4svR(-) Tumor	1.01 ± 0.30	0.29 ± 0.20	0.17 ± 0.02	0.18 ± 0.05	0.11 ± 0.04

Table S10. Biodistribution of [¹⁷⁷Lu]Lu-GAS3 in male SCID mice bearing twin HEK293-CCK2i4svR and wtHEK293 tumors at 1, 4, 24, 48, 72 and 96 h pi, expressed as %IA/g and representing mean values ± SD, n=4.

Organs	%IA/g ± SD, n=4				
	4 h	24 h	48 h	72 h	96 h
Blood	3.19 ± 0.44	0.07 ± 0.01	0.05 ± 0.01	0.04 ± 0.00	0.04 ± 0.00
Liver	2.73 ± 0.31	0.51 ± 0.05	0.35 ± 0.09	0.32 ± 0.02	0.25 ± 0.02
Heart	0.94 ± 0.21	0.08 ± 0.02	0.08 ± 0.01	0.04 ± 0.01	0.03 ± 0.01
Kidneys	8.86 ± 0.67	1.42 ± 0.29	0.47 ± 0.07	0.17 ± 0.04	0.11 ± 0.03
Stomach	0.64 ± 0.07	0.11 ± 0.01	0.08 ± 0.01	0.06 ± 0.02	0.04 ± 0.00
Intestines	1.85 ± 0.18	0.17 ± 0.04	0.07 ± 0.01	0.04 ± 0.01	0.03 ± 0.01
Spleen	1.19 ± 0.12	0.45 ± 0.05	0.43 ± 0.10	0.44 ± 0.07	0.39 ± 0.02
Muscle	0.31 ± 0.04	0.03 ± 0.01	0.04 ± 0.01	0.03 ± 0.01	0.01 ± 0.00
Lungs	1.94 ± 0.21	0.26 ± 0.02	0.18 ± 0.03	0.15 ± 0.01	0.14 ± 0.01
Femur	0.60 ± 0.07	0.11 ± 0.02	0.14 ± 0.04	0.06 ± 0.00	0.06 ± 0.00
Pancreas	1.00 ± 0.13	0.07 ± 0.01	0.08 ± 0.04	0.03 ± 0.01	0.02 ± 0.00
CCK2i4svR(+) Tumor	20.88 ± 1.20	5.38 ± 0.77	3.39 ± 0.23	1.99 ± 0.31	1.52 ± 0.43
CCK2i4svR(-) Tumor	1.14 ± 0.50	0.37 ± 0.12	0.25 ± 0.14	0.13 ± 0.03	0.00 ± 0.04

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